



Mitochondrial function maintenance and mitochondrial training in ageing and related diseases

Gan Gao^a, Zhihui Xie^b, Hongliang Huang^{a,c,d,*}

^a School of Pharmacy, Guangdong Pharmaceutical University, Guangzhou, 510006, China

^b Xie Zhihui Biomedical Research Institute Guangzhou Co. Ltd., Guangzhou, 510006, China

^c School of Biosciences and Biopharmaceutics, Guangdong Pharmaceutical University, Guangzhou, 510006, China

^d Key Laboratory of New Drug Discovery and Evaluation, Guangdong Pharmaceutical University, Guangzhou, 510006, China

ARTICLE INFO

Keywords:

Cellular senescence

Mitochondrial quality control

Anti-ageing

Age-related diseases

ABSTRACT

Cellular senescence driven by mitochondrial dysfunction is a key contributor to ageing and related diseases. The decline in the quality and quantity of healthy mitochondria with ageing disrupts energy production, redox homeostasis, and intracellular signaling. Mitochondrial quality control (MQC) is a cellular self-repair mechanism that protects mitochondrial function and maintains a healthy mitochondrial network. Targeted regulation of MQC is expected to moderate the development of cellular senescence and related diseases. We explored the impact of mitochondrial function on cell fate at the molecular and organelle levels, analyzed the role of mitochondria-targeted interventions for delaying cellular senescence and ameliorating age-related diseases, and pointed out the idea of increasing the critical level of healthy mitochondria to cope with internal and external stressful stimuli and to improve the ability of self-repairing by exercising and protecting mitochondria in the long term.

1. Introduction

The increase in the aged population brings more challenges than ever. The human lifespan has been extended considerably with the advancement of science and technology, especially in healthcare. Ageing has become a significant risk factor for cancer, cardiovascular disease, and neurodegenerative diseases. The World Health Organization (WHO) defines senescence as the gradual accumulation of molecular and cellular damage in the body over time. Molecular and cellular damage associated with senescence is the primary cause of ageing.¹ It is well known that ageing is closely related to various diseases. The function of the significant organs gradually deteriorates in older people, and they gradually suffer various age-related diseases.

Mitochondria are the leading energy suppliers in eukaryotic cells, and lots of studies have illustrated that their functional degradation is associated with the occurrence and development of age-related diseases.² The oxidative metabolism of the mitochondria provides energy for the operation of cellular metabolism; the mitochondria themselves are also constantly subject to oxidative damage. As the damage accumulates, the function of mitochondria gradually weakens, leading to more dysfunction of mitochondria, which can accelerate cell ageing. Mitochondria are

essential for cellular physiological activities and signaling and may also be an initiating factor for cell damage or death.³ Without external stressors, the cell maintains mitochondrial structure and function stability through MQC (Mitochondrial Quality Control). This process ensures mitochondria's stable and efficient function and prevents cellular dysfunction and related diseases caused by damage to mitochondria.⁴ It has been demonstrated that a certain degree of enhancement of MQC can help prevent the development of various diseases, particularly those associated with ageing. However, excessive activation of the control pathway can also result in cellular damage and reduced mitochondrial function.⁵ Healthy ageing can be achieved through moderate exercise, a proper diet, and targeted mitochondrial repair and functional activation (Fig. 1). Specific intervention strategies are employed to enhance cellular mitochondria quantity, quality, and functionality. This is intended to optimize cellular energy production and utilization, enhance metabolic capacity, and improve overall performance. Mitochondrial training may be defined as a process of continuous activation of mitochondrial function and quality control, with the increase of the capacity of response to cellular stress for mitochondria.

Meanwhile, it is known that mitochondrial DNA mutated more with ageing, but low levels of mutated mitochondrial DNA did not affect mitochondrial function or normal cellular energy metabolism. Usually, a

* Corresponding author. School of Pharmacy, Guangdong Pharmaceutical University, Guangzhou, 510006, China.

E-mail address: huanghl@gdpu.edu.cn (H. Huang).

Peer review under the responsibility of Editorial Board of Journal of Holistic Integrative Pharmacy.

<https://doi.org/10.1016/j.jhip.2025.04.001>

Received 19 November 2024; Received in revised form 20 March 2025; Accepted 23 April 2025

2707-3688/© 2025 The Authors. Publishing services by Elsevier B.V. on behalf of KeAi Communications Co. Ltd. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

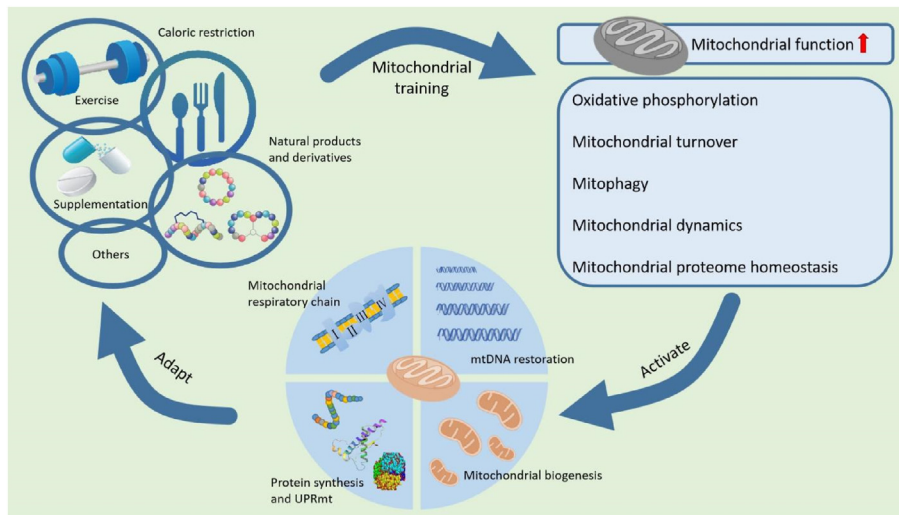


Fig. 1. Mitochondrial training. Strategies focusing on improving mitochondrial function have been paid more attention. For example, caloric restriction (CR) can reduce oxidative damage in the body. Endurance exercise training enhances the aerobic capacity of skeletal muscle by improving the quality control, content, and function of mitochondria. SIEGEL M P et al. showed that a single treatment with a mitochondrial-targeting peptide can rapidly restore mitochondrial energy to youthful levels in elderly mice, reversing declines in maximal mitochondrial ATP production, oxidative phosphorylation coupling, and cellular energy supply. These changes are more pronounced in ageing animals.^{9–11} Mitochondrial training refers to an approach that involves sustained interventions such as exercise, dietary modifications, natural active products, or lifestyle modifications to achieve rapid cellular adaptation and sustained resistance to stress responses and pressures by enhancing MQC and optimizing energy production. This process would be a potential intervention to slow cellular senescence, reduce age-related dysfunction, and promote healthy ageing.

phenotypic manifestation of the genetic defect occurs only when a threshold level is exceeded.⁶ Thus, within the threshold range, the cell maintains a normal mitochondrial network state by activating mitochondrial quality control. This is evidenced by the increased number of healthy mitochondria, protection of mitochondrial DNA integrity, and maintenance of proteome and active respiratory chain complex reserves.⁷ In this way, the training of mitochondria may have a more vital function and tolerance against cellular ageing in the face of progressively deteriorating conditions.

The 12 hallmarks of ageing include genomic instability, telomere attrition, epigenetic alterations, loss of proteostasis, disabled macroautophagy, deregulated nutrient sensing, mitochondrial dysfunction, cellular senescence, stem cell exhaustion, altered intercellular communication, chronic inflammation, and dysbiosis.⁸ The inherent fragmentation of the markers is counterbalanced by their interdependence, with the influence of one marker on the others. It is therefore recommended that each feature be considered as an entry point for future exploration of the ageing process, as well as the development of new anti-ageing drugs. Next, we will focus on the relationship between mitochondrial function and cellular senescence, illustrate the potential relationship between mitochondrial dysfunction and ageing, and provide an overview of emerging anti-ageing strategies targeting mitochondria.

2. Cellular senescence and related diseases

Cellular senescence was first discovered in human diploid cell lines; these cells go through active multiplication, gradual cessation of mitotic

activity, and accumulation of cellular depreservation activation of the cyclin-dependent kinase (CDK)-inhibitor 1 (CIP1), causing cell growth to stall.^{12,13} This phenomenon typically contributes to the maintenance of genetic stability. It can slow or inhibit the transformation of preneoplastic lesions into neoplastic lesions during tumorigenesis, suggesting that normal cell senescence can inhibit cancer and tumorigenesis but is reversed by irreversible functional impairment and overaccumulation. Cell senescence is often accompanied by the development of senescence-associated secretory phenotype (SASP); SASP consists of pro-inflammatory factors, growth factors, chemokines, and matrix remodelling enzymes. They can cause low-grade chronic inflammation and diseases in the body and accelerate the senescence of cells or neighbouring cells (Fig. 2).^{14,15}

2.1. Cardiovascular system and cellular senescence

The heart is an essential organ of the human body. Still, with the increase of age, the gradual increase of senescent cells will harm the myocardial structure and function, which leads to the decline of myocardial contractility and diastolic function and the decline of the heart's pumping ability, eventually causing heart failure. Cardiac ageing is a heterogeneous process characterized by increased reactive oxygen species (ROS), genomic DNA damage, and telomere shortening. Supraphysiological concentrations of ROS oxidatively modify biological macromolecules, including proteins, nucleic acids, and lipids. This process generates harmful substances and oxidative damage to cellular organelles, impairing cellular function and contributing to gradual ageing.

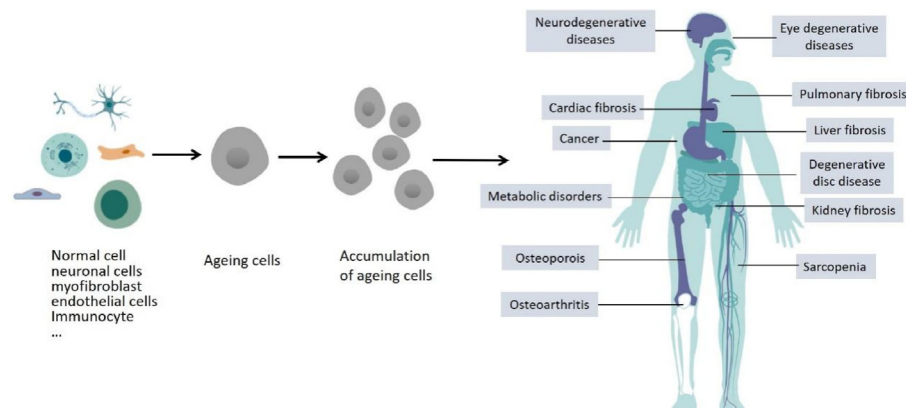


Fig. 2. Cellular senescence is associated with diseases of senescence. DNA damage, protein misfolding, mitochondrial dysfunction, and other adverse phenomena inevitably occur in cells. Normal cells activate self-protective mechanisms such as DNA repair, autophagy, antioxidative stress, and mitochondrial fusion and division to combat these adverse factors. However, cellular senescence with age is a response to this imbalance of damage and repair, which has been linked to various age-related diseases, including neurodegenerative diseases, metabolic dysfunction, and age-related diseases.

Normal cells can resist oxidative damage caused by this ROS overload through various enzymes and signaling pathways of the antioxidant system. However, senescent cells exhibit an imbalance in the antioxidant system, ultimately leading to the ageing or death of cells, known as oxidative stress (OS).¹⁶ With cellular senescence, particularly of the vascular endothelium, functional changes in morphology (e.g., flattened endothelial cells) occur, which lead to endothelial dysfunction, atherosclerosis, and ultimately hypertension. Bone marrow-derived endothelial progenitor cells and klotho are reduced during endothelial cell senescence.¹⁷ Some studies have shown that ageing increases ROS accumulation and oxidative stress in the heart.^{18–20} The non-selective damage induced by oxidative stress has been strongly associated with a variety of age-related cardiovascular diseases, including atherosclerosis, myocardial infarction, and heart failure.^{21,22} This is because oxidative stress increases the number of oxidized macromolecules and the level of ROS production.²³ At the same time, mitochondria are essential for endogenous ROS production. During physiological ageing, mitochondrial membrane potential (MMP) decreases, resulting in reduced energy production and increased OS due to the production of ROS.^{24,25} The accumulation of dysfunctional mitochondria within cells can lead to various cardiovascular diseases associated with ageing.²⁶

Consequently, both antioxidants and mitochondrial protection represent significant therapeutic targets for improving age-related cardiac phenotypes.^{27–29} Another factor contributing to the deterioration of heart function is the vicious cycle of the inflammatory response of ageing cells and oxidative stress damage. Biomarkers of inflammation are consistently linked to chronic disease risk. For example, high levels of interleukin-6 (IL-6) and tumor necrosis factor- α (TNF- α) are associated with an increased risk of cardiovascular disease.³⁰ Inflammation caused by cell death will produce excessive ROS, leading to oxidative stress damage, which induces inflammation, forming a vicious cycle in ageing cardiomyocytes.^{31,32} Based on the unique features of senescent cells, current efforts are primarily invested in discovering methods to eliminate senescent cells. Pan-tyrosine kinase inhibitor Dasatinib and a natural flavonoid, quercetin (D/Q combination), have been shown to kill senescent human umbilical vein endothelial cells (HUEVCs) in tissue culture selectively and to suppress the senescent cell signature under a set of pathological conditions.³³ R. Schmitt et al. designed siRNA or the small molecule ABT-737 to target and inhibit the expression of the anti-apoptotic proteins BCL-W and BCL-XL in senescent cells, specifically triggering the death of senescent cells and inhibiting the adverse cardiovascular effects of biological ageing.³⁴ In addition to removing senescent cells, another promising tool for targeting senescent cells is eliminating SASP without causing cell death. For example, metformin has been shown to improve cardiovascular function and prolong lifespan in diabetic patients, providing support for investigation into the ability of metformin to delay ageing in nondiabetic persons.³⁵ Depending on cell type, the senescence stage, and the senescence inducer type, the SASP is highly heterogeneous, and it is difficult to identify a general regulatory mechanism.³⁶ Therefore, there is a need to analyze individual cells by single-cell RNA sequencing to understand better the specific SASP components that drive specific functions in cardiovascular ageing. More clinical research is urgently needed. Senescent cells accumulate in the heart during ageing, resulting in decreased cardiac function, heart damage, and an increased risk of heart disease. OS and mitochondria are two key targets that require further investigation.

2.2. Nervous system and cellular senescence

Similarly, in aged people, the degeneration of the central nervous system (CNS) drives the progressive impairment of cognitive and physical abilities, the ageing brain being vulnerable to Alzheimer's disease (AD), Parkinson's disease (PD), and stroke.^{37,38} One in 10 individuals aged 65 or older suffers from AD, and its prevalence continues to increase with ageing.³⁹ Cellular ageing increases in AD, PD, Amyotrophic Lateral Sclerosis (ALS), and Huntington's disease (HD), including microglia,

astrocytes, and neuronal cell senescence. A growing body of evidence indicates that oxidative stress accelerates brain ageing and related diseases. Furthermore, it suggests that free radicals damage mitochondria and increase the production of toxic β -amyloid protein (A β), which is thought to be a significant cause of AD.⁴⁰ Neurons rely on the energy produced by mitochondria to maintain synapses and neurotransmission. Consequently, brain mitochondria are more active than other tissues.⁴¹

Nevertheless, cells within the AD brain are susceptible to oxidative damage and exhibit reduced mitochondrial respiration, reduced movement, reduced size, and disruption of the inner membrane of the cristae relative to normal brain cells.⁴² Decreased endogenous antioxidant production in the ageing brain⁴³; the redox environment disturbance triggers conformational changes and mitochondrial localization of the pro-apoptotic BCL-2 family (BAX, BAK, BAD, BIM). Furthermore, it has been demonstrated that increased mitochondrial membrane permeability leads to the release of pro-apoptotic proteins from the mitochondrial membrane space, including cytochrome c; this process activates the cysteinyl aspartate specific proteinase (Caspase-3), which initiates apoptosis by cleaving numerous essential cellular proteins through proteolytic hydrolysis.^{44,45} This is why severe nigrostriatal dopamine neuron loss occurs in PD. Therefore, antioxidant supplementation and mitochondrial protection prevent ageing and ameliorate pathological changes in age-related brain diseases. In recent years, inflammation has also been identified as an essential contributor to neurodegenerative diseases. Inflammatory responses can clear toxins and unknown pathogens and promote cytotoxicity and neurodegeneration.⁴⁶ However, it is unclear whether inflammation is a consequence or a positive factor in neurodegenerative diseases. In particular, the regulatory function of mitochondria between oxidative stress and inflammatory responses in brain cells.⁴⁷ Mitochondrial dysfunction leads to excess ROS production, which brings about oxidative damage to glial cells, and normal cells activate the autophagy pathway to remove damaged mitochondria. While senescent cells are defective in autophagy, leading to the accumulation of damaged mitochondria that release their components into the cell, mtDNA, in particular, can act as a damage-associated molecular pattern (DAMP), which is sensed as a non-self molecule and triggers an innate immune-inflammatory response by binding to danger signaling receptors.⁴⁸ Microglia are significant players in neuroinflammation, and overactivated microglia release pro-inflammatory factors in response to abnormal accumulation of disease-causing proteins and invasion by pathogens.⁴⁹ Uncontrolled neuroinflammation can induce chronic inflammatory responses and accelerate the progression of neurodegenerative diseases such as AD.⁵⁰ In mouse models of neurodegenerative diseases, eliminating senescent cells improves brain function and alleviates cognitive dysfunction during ageing.⁵¹

2.3. Skeletal muscle system and cellular senescence

Bone and skeletal muscle are involved in a variety of physiological activities such as locomotion, protection, metabolism, and maintenance of the systemic microenvironment, and their structure and function are affected by endocrine, inflammatory, and external stresses (physical activity, drugs, pollution, and diet).⁵² Disrupting the metabolic balance between anabolism and catabolism in ageing bone and muscle results in tissue damage characterized by disruption of the skeletal microenvironment and accumulation of senescent cells in muscle.⁵³ Skeletal muscle represents a significant site of energy production and expenditure. The presence of numerous and active mitochondria contributes to the maintenance of energy metabolic homeostasis.⁵⁴ However, the accumulation of age-related oxidative stress and DNA damage above the threshold for selective initiation of senescence in normal cells resists endogenous stresses through apoptosis. This also means it is difficult to cope with exogenous stresses that result in irreparable tissue damage.⁵⁵ Even in cellular senescence, the inability to remove these cells promptly due to a compromised immune system can also result in the accumulation of senescent cells with a reduced regenerative capacity, contributing to the

gradual ageing of the tissues.⁴⁶ Diseases of the immune system caused by senescence of immune cells will be discussed later. Sarcopenia, which reduces the quality of life in the old and causes them to be more prone to falls and fractures, is characterized by type II fibre atrophy, changes in fibre size and heterogeneity, accumulation of fat and connective tissue between fibres, decreased mitochondrial metabolism and its oxidative capacity, inflammation, and increased protein catabolism.⁵⁶ Bones have a vast reservoir of progenitor cells in the body. It has been shown that various cell types in the bone microenvironment are senescent during ageing and form heterogeneous SASPs that affect normal osteoblasts.⁵⁷ Age-related bone loss is associated with the accumulation of senescent cells, particularly osteoblasts, in the bones of older individuals. As mentioned, many senescent skeletal muscle cells are involved in age-related skeletal diseases. Removing senescent skeletal muscle cells from the skeletal system is critical for slowing skeletal ageing and alleviating disease. In INK-ATTAC or p16-3MR transgenic mice, drugs targeting *p16Ink4a* successfully remove *p16Ink4a*-positive senescent skeletal muscle cells, alleviating age-related bone loss and extending the lifespan of mice.^{58,59} Targeting cellular senescence has led to the emergence of "Senolytics" (selectively eliminating senescent skeletal muscle cells) and "Senomorphics" (inhibition of SASP secretion) to neutralize and eliminate the adverse effects of senescent skeletal muscle cells, which are collectively referred to as "chemotherapeutic" strategies.⁶⁰

Furthermore, removing senescent cells is not optimal; inhibiting cellular ageing and alleviating senescence-induced cellular damage are more beneficial. Oxidative stress and inflammation are significant factors that are closely associated with the process of ageing. Excessive ROS production in senescent cells leads to mitochondrial dysfunction and further exacerbates inflammation.⁶¹ Mitochondria are both the primary site of ROS production and the target of ROS.⁶² The synthesis and breakdown of proteins in bone and muscle require much energy, leading to the mitochondria's continuous production of loaded ROS. With the age-related decline in cellular autophagy,^{63,64} damaged mitochondria cannot be removed promptly and are not replenished by healthy mitochondria, which reduces ATP synthesis. This, in turn, is associated with a reduction in protein synthesis, which ultimately causes tissue damage.⁶⁵ Furthermore, oxidative damage to the mitochondrial membrane results in the release of intra-mitochondrial components which, in turn, initiates an immune-inflammatory response and mitochondria-mediated apoptosis.⁶⁶

2.4. Other body systems and cellular senescence

2.4.1. Liver, lungs and kidney

The liver is the most significant metabolic organ. Almost all the substances absorbed from the gastrointestinal tract are transported to the liver, where they undergo synthesis, degradation, and storage transformation. Therefore, age-related changes in liver function increase the body's susceptibility to age-related diseases.⁶⁷ Therefore, age-related changes in liver function increase the body's susceptibility to age-related diseases.⁶⁸ Hepatic parenchymal cells are the liver's primary cells; with ageing, they accumulate lipids, have mitochondrial dysfunction, and increase ROS production.⁶⁹ This is the main reason for the high prevalence of fatty liver in older people.⁷⁰ Kupffer cells are resident hepatic macrophages located in the liver sinusoids and involved in the innate immune response to liver infection.⁷¹ The effects of ageing on macrophages are a decrease in phagocytosis and autophagy, as well as an increase in the production of cytokines that contribute to the inflammatory phenotype ('inflammaging') of old age.⁷¹ Examples include TNF- α and IL-6. Reduced oxidative phosphorylation capacity, decreased membrane potential, increased mtDNA mutations, OS, and morphological changes have been reported in the mitochondria of ageing livers compared to that of the young state.⁷² This is associated with the production of supraphysiological concentrations of ROS, while mitochondrial dysfunction promotes an increase in ROS, which also drives cellular senescence.⁷³ Liver fibrosis progresses to cirrhosis, an advanced form in

which damaged tissues and cells cannot be repaired due to large amounts of extracellular matrix and the restoration of normal liver structure, marking the end of liver function and complete liver failure, and ultimately death. The most important factors are ROS and cellular senescence. The presence of senescent cells disrupts tissue structure and function and increases the production of senescent cells in liver tissue, contributing to fibre formation.⁷⁴ It is worth mentioning that fibrosis is a typical ageing change and is common in the tissues of the ageing organism. For example, prostate fibrosis in older men leads to urinary tract dysfunction (UTD), in whose tissues a decrease in mitochondrial function and an increase in cellular senescence are observed.⁷⁵

Young normal lungs in which a small number of senescent cells are rapidly cleared by immune cells and normal cells maintain a baseline of telomere length and mitochondrial homeostasis. As fibrosis increases and the number of senescent cells increases, the immune cell response to senescent cell clearance becomes slow. Senescent cells are characterized by telomere shortening, secretion of high rates of SASP, mitochondrial dysfunction, and imbalances in mitochondrial division and fusion.⁷⁶ Chronic Obstructive Pulmonary disease (COPD) is a major healthcare problem with high morbidity and mortality. Although there is no known cure, COPD is managed through lifestyle changes and medication, but these therapies have limitations and new treatments need to be explored. Studies have shown increased expression of SA- β gal, p16, and p21 in endothelial colony-forming cells (ECFCs) derived from COPD patients, demonstrating an increase in senescent cells.⁷⁷ Pulmonary fibrosis is a significant category of end-stage pathological changes of lung diseases characterized by fibroblast proliferation and accumulation of many extracellular matrices accompanied by inflammatory damage and tissue structure destruction. For example, normal alveolar tissue is damaged, and scarring forms. It has been reported that the disease Idiopathic Pulmonary Fibrosis (IPF) is mediated, in part, by senescent cells, and cellular senescence markers are detectable within IPF lung tissue, which can be targeted to restore healthy pulmonary tissue of ageing mice.⁷⁸ For example, upregulation of senescence-related pathways in type II alveolar epithelial cells (AT II) has been noted in a mouse model of IPF where AT II Sin3a has been knocked out to induce senescence.⁷⁹

Halloran and colleagues first discussed that the accumulation of senescent cells may be responsible for the insufficient repair capacity and functional loss in older kidneys almost 20 years ago.^{80,81} During ageing, intrinsic cellular responses that repeat minor injuries continue to occur, and these injuries go virtually unnoticed for a long period until a certain threshold is reached that can impact kidney function.⁵⁸ Renal ageing and Chronic Kidney disease (CKD) share many common features, ranging from pathological and clinical manifestations to underlying mechanisms. Mechanisms common to the renal ageing process and the development of CKD include increased cellular senescence, decreased autophagy, mitochondrial dysfunction, and altered epigenetic regulation, suggesting that cellular senescence would be a potential therapeutic target applicable to both diseases.⁸² Early on, for end-stage renal disease, renal replacement therapy was required, and with the development of key mechanisms of senescence such as cell cycle blockade, apoptosis inhibition, and SASPs, selective targeting of senescent cells (adopting Senolytics therapy) or systemic SASPs (adopting Senomorphics therapy) has been progressively investigated and applied in basic models of CKD and clinical trials.⁸³ In a study, Peter de Keizer and colleagues identify forkhead box O 4 (FOXO4) as a key regulator of senescent cell viability and demonstrate that targeting FOXO4 signaling causes apoptosis and restores renal function in rapidly ageing and naturally aged mice.⁸⁴ Meanwhile, SASP can accelerate the renal ageing process in different environments and participate in the pathological process of renal diseases, and therapeutic strategies targeting this have potential.

2.4.2. Skin and eyes

The skin consists of different cell types with different proliferative capacities. During the ageing process, senescent cells accumulate in different compartments of the human skin, leading to impaired skin

physiology. Skin ageing is characterized by slow regeneration and even permanent erosion of skin structure and function. Skin ageing is associated with impaired protection, in particular impaired wound healing and barrier function, increased inflammation, impaired water and thermal homeostasis of organisms, and susceptibility to various skin diseases, including cancer.⁸⁵ Senescent cells accumulate as the skin ages, leading to a decline in all aspects of skin function. Paracrine signaling between dermal fibroblasts and keratin-forming cells is necessary for tissue homeostasis in physiological conditions and skin diseases.⁸⁶ The crosstalk between these cell populations is altered during the ageing process.⁸⁷

Ageing leads to a gradual decline of function in multiple organs. Cataracts, glaucoma, diabetic retinopathy, and age-related macular degeneration (AMD) are age-related ocular diseases. In the old human retina, p16INK4a is expressed in rods, ganglion cells, amacrine cells, and horizontal cells. Moreover, p16INK4a and p21Cip1 are expressed in retinal vascular vessels, elucidating the expression of canonical senescence markers in retinal cells.⁸⁸ In recent years, glaucoma has emerged as a blinding ocular disease intricately linked to cellular senescence. The principal pathways implicated are oxidative stress, mitochondrial dysfunction, DNA damage, autophagy impairment, and the secretion of various senescence-associated secretory phenotype factors.⁸⁹ AMD is a complex disease that is closely related to cellular senescence. Senescent ocular cells exhibit mitochondrial and telomere dysfunction as well as elevated oxidative stress, leading to the production of SASP and more ROS, both of which can induce and stabilise senescence in neighbouring cells.⁹⁰

2.4.3. Metabolic systems

With the increase of age, the elderly group has hyperglycemia, fatty liver, hyperlipidemia, hypertension, high uric acid, and other diseases. A disease called "metabolic syndrome" disturbs the body's metabolism and damages human health. Ageing is thought to be a risk factor for metabolic syndrome, especially in obese older adults. Senescent cells accumulate in the adipose tissue of obese people and rodents. Experiments have shown that targeted elimination of these senescent cells can reduce the burden of senescent cells in obese mice, alleviate metabolic dysfunction and adipose tissue inflammation, improve glucose tolerance, enhance insulin sensitivity, and reduce the release of inflammatory mediators in obese mice. At the same time, senescent cells can cause macrophage migration into adipose tissue in obesity, and targeting senescent cells prevents and reduces the adipose tissue macrophage accumulation often associated with obesity.⁹¹

2.4.4. Immune system

As the human organism ages, immune cells undergo a process of gradual senescence, leading to a decline in immune system function and, in turn, an increased susceptibility to a variety of immune-related diseases.⁹² T cells, as a core immune system component, undergo significant changes during the ageing process.⁹³ The initial T-cell pool becomes smaller, and the memory T-cell pool becomes larger, decreasing the number of available TCRs. The senescent T cells exhibit molecular features such as mitochondrial dysfunction, epigenetic remodelling, DNA damage, and short telomeres.⁹⁴ These senescent T cells secrete pro-inflammatory factors that trigger chronic inflammation, disrupting homeostasis in tissues and causing cardiovascular diseases, metabolic disorders, and neurodegenerative diseases.^{95,96} Recently, interventions targeting senescent cells have become a hot research topic. Removing senescent cells or regulating their function is expected to slow down the ageing process and improve the function of the immune system.

The administration of Senolytics drugs, which induce apoptosis, eliminates senescent cells. For instance, the combination of Dasatinib and quercetin effectively removes a wide range of senescent cells.⁹⁷ In addition, cardiac glycosides have been shown to induce senescent cell death by altering the inner membrane potential and proton concentration of senescent cells. Modulation of the immune system enhances the body's capacity to clear senescent cells. For instance, CAR-T cell therapy can

specifically recognize and remove senescent cells.⁹⁸

Furthermore, gene editing technology has emerged as a potent tool for targeting key genes in senescent cells, impeding their senescence process. A study has shown that the accumulation of senescent cells can be reduced by inhibiting the *p16INK4a* gene.⁹⁹ Immune cell senescence has been identified as a significant factor in developing immune system diseases. Interventions that target senescent cells have emerged as a novel approach for delaying senescence and enhancing immune system function.¹⁰⁰ It is recommended that future studies further explore the mechanism of action and the prospect of clinical application of these interventions, which can provide a theoretical basis for the development of effective anti-ageing therapies.

3. Mitochondrial dysfunction promotes cellular senescence

Complex life is composed of eukaryotic cells, and the presence of mitochondria in eukaryotic cells is essential to evolution.^{101,102} Mitochondria are organelles with a double membrane structure composed of an inner mitochondrial membrane (IMM) and an outer mitochondrial membrane (OMM). The IMM folds inward to form cristae, which provide more attachment sites for enzymes and generate energy to participate in various intracellular reactions through aerobic respiration. They are usually filamentous or rod-shaped slender bodies, and the mitochondria have a large matrix volume, making part of the gap between the IMM and the OMM very small, which is called the mitochondrial inner boundary membrane (IBM).^{103,104} Despite this diversity, all mitochondria derive from a common ancestral organelle that originated from integrating an endosymbiotic alphaproteobacterium into a host cell related to Asgard archaea.¹⁰⁵ The essential function of mitochondria is to form an electrochemical gradient on the respiratory chain of the inner membrane through the coupled transfer of electrons to oxygen and the transport of protons from the matrix through the IMM into the intermembrane space, which provides power for the terminal complex V of the respiratory chain and catalyzes the synthesis of ATP in most cells under the action of ATP synthase. Electrochemical gradients are also used for other critical mitochondrial functions, such as maintaining homeostasis for transporting buffer signaling ions Ca^{2+} .^{106,107} Reducing the electrochemical gradient signals cell activation to repair and eliminate defective mitochondrial pathways.¹⁰⁸

It was earlier found in the ageing process of model animals that male houseflies enter the old age stage during the second week of adult life, as reflected in the loss of wings, and the activity of magnesium-activated adenosine triphosphatase in the giant mitochondria of the flight muscle decreases with failure in flight. The rapid decline in the activity of the α -glycerophosphate dehydrogenase dependent on nicotinamide adenine dinucleotide is a sign of wing loss in ageing male houseflies.⁴⁶ Type 2 diabetes mellitus (T2DM) is the most common chronic metabolic disease in the ageing population and is characterized by impaired glucose tolerance due to insulin-stimulated glucose metabolism dysfunction in the skeletal muscle.^{105,109–111} IR in older adults is associated with increased intracellular fatty acid metabolites, possibly due to age-related decreased mitochondrial oxidation and phosphorylation activity.¹¹² In conclusion, the ageing process is accompanied by a decrease in mitochondria-related enzyme activities and a drop in mitochondrial quality. For example, citrate synthase activity declined, mitochondria's respiratory capacity attenuated, ROS production rose, membrane permeability increased, and damaged protein aggregation and mitophagy levels were reduced.¹¹³ So, what is the link between mitochondrial dysfunction and cellular ageing? This is discussed next.

3.1. Mitochondrial biogenesis

The cells launch the mitochondrial biogenesis process in response to the energy demand triggered by developmental signals and environmental stressors. This is a self-renewal route by which new mitochondria are generated from existing ones, and the process of mitochondrial

biogenesis occurs mainly in healthy cells.¹¹⁴ (Fig. 3). If mitochondrial biogenesis is attenuated, mitochondrial renewal slows down, and overall productivity decreases, resulting in an increased accumulation of damaged biomacromolecules, further exacerbating mitochondrial dysfunction in ageing cells.^{115,116} In ageing, the imbalance between the production of healthy mitochondria and the clearance of abnormal mitochondria accounts for the dysfunction of mitochondrial renewal and the accumulation of damaged mitochondria in cells. Usually, cells undergo mitophagy to degrade damaged mitochondria while simultaneously initiating compensatory biosynthetic reactions to replenish their mitochondrial pool in response to a deficiency in biological energy and reduced ATP synthesis. Mitochondria are the energy basis for the activity of neurons in the human brain. Activating mitochondrial biogenesis is beneficial in delaying the onset and development of neurodegenerative diseases in the elderly.^{117–119} Mitochondrial dysfunction due to the reduction of biogenesis in the blood is a potential marker to assess the risk of PD.⁴⁷ The peroxisome proliferator-activated receptor gamma coactivator-1 α (PGC-1 α) modulates mitochondrial biogenesis. More expression of PGC-1 α and the adenosine 5'-monophosphate (AMP)-activated protein kinase (AMPK) have been thought of as a method to extend the healthy life of the elderly.¹²⁰

PGC-1 α is a significant regulator of mitochondrial biogenesis and

function.^{121,122} PGC-1 α regulates the expression of mitochondria-associated genes by interacting with multiple transcription factors. On the one hand, PGC-1 α can activate the expression of NRF1, which further promotes the expression of mitochondrial DNA polymerase γ (POLG) and mitochondrial transcription factor A (TFAM), and TFAM acts on the nuclear genome to initiate mitochondrial DNA replication and transcription. On the other hand, PGC-1 α can bind to estrogen-related receptor α (ERR α) to form a complex that enhances the transcriptional activity of ERR α . ERR α activates the expression of a range of downstream genes that promote mitochondrial biogenesis.⁶³ When PGC-1 α is overexpressed, mitochondrial biogenesis is activated, while PGC-1 α expression is inhibited, and mitochondrial biogenesis activity is decreased.^{123,124} It turns out that AMPK is the upstream activator of PGC-1 α , and AMPK enhances mitochondrial biogenesis not only by inducing *PGC-1 α* transcription but also by activating PGC-1 α through phosphorylation of threonine-177 and serine-538.¹²⁵ Simultaneously, the silent information regulator 1 (*SIRT1*) can directly act on PGC-1 α to mediate its deacetylation and improve the activity of PGC-1 α to enhance mitochondrial biogenesis, especially in the case of caloric restriction (CR).¹²⁶ Hydrogen sulfide is an essential regulator of cardiac mitochondrial content. Exogenous hydrogen sulfide can induce mitochondrial biogenesis through the AMPK-PGC-1 α signaling cascade, improve mitochondrial respiration, increase ATP production efficiency,

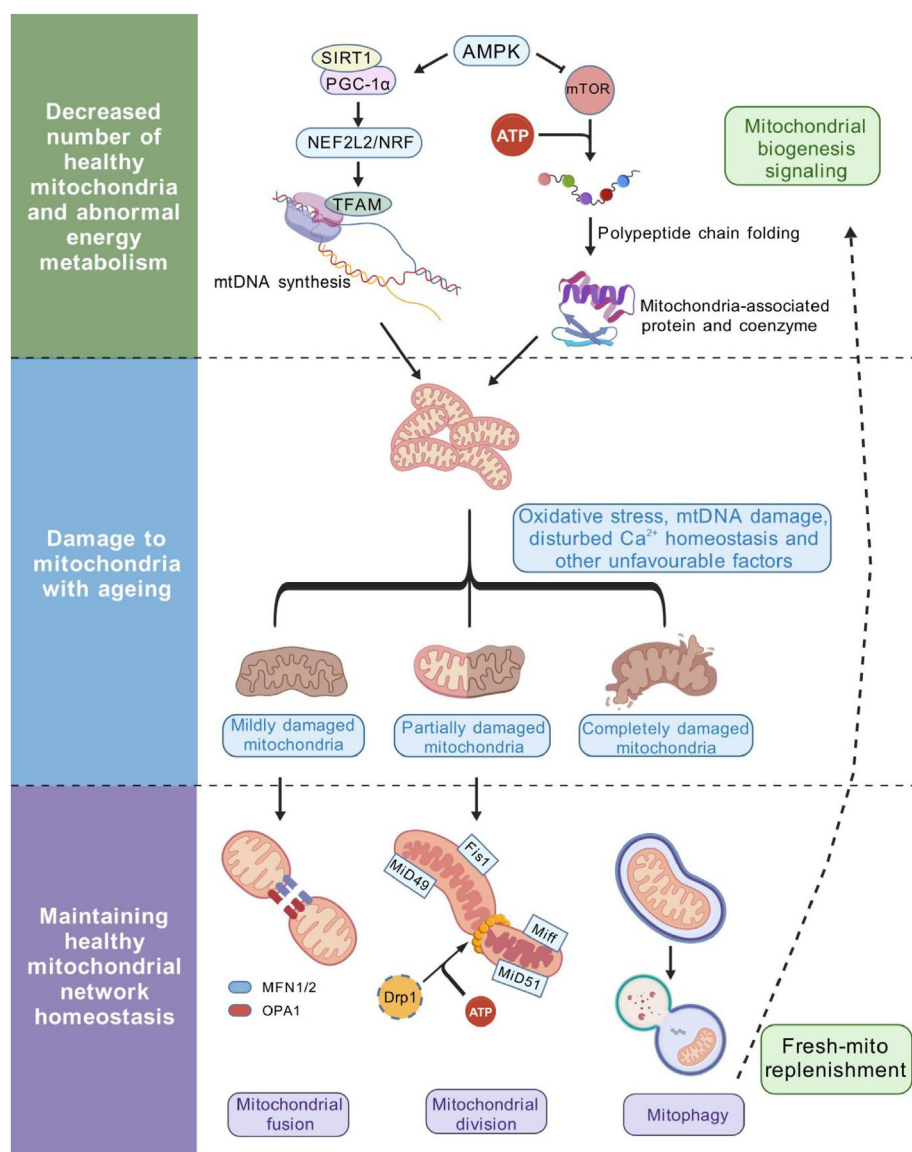


Fig. 3. Mitochondrial network homeostasis. The decrease in ATP synthesis activates the AMPK signaling pathway, then AMPK induces PGC1- α to act on the transcription factor NEF2L2. NEF2L2 acts on mitochondrial transcription factor (TFAM) to drive mtDNA transcription and translation. SIRT1 can directly mediate PGC1- α deacetylation to activate it. On the other hand, AMPK inhibits mTOR, induces the synthesis of protein, and promotes healthy mitochondrial biogenesis. Mitochondrial fusion was regulated by OPA1, MFN1, and MFN2 and finally formed a complete mitochondrial reticular state. If mitochondria are damaged, division occurs, and endoplasmic reticulum-derived membrane structures mark the division site. The OMM proteins MID49, MID51, and FIS1 recruit DRP1, which binds to MFF. The dimers and oligomers of DRP1 assemble into a helical superstructure at the division site. Shrinks hydrolysis to cut mitochondria, and the damaged mitochondria are partially cleared by the mitochondrial autophagy pathway. Then, the decline in mitochondrial numbers stimulates new mitochondrial biogenesis.

and improve cardiac function.¹²⁷ Transcription factor NF-E2 p45-related factor 2 (NRF2) allows cells to adapt and survive under stressful conditions by regulating the gene expression of various cellular protective protein networks. It activates mitochondrial biogenesis by up-regulating the nuclear respiratory factor 1 and PGC-1 α .¹²⁸

3.2. Mitophagy

Autophagy refers to the ability of cells to provide nutrients and energy by degrading non-essential components in response to the transient stress of survival, thus sustaining life. Correspondingly, autophagy may degrade potentially toxic proteins to prevent cell damage or the process of apoptosis. Autophagy degrades components in the cytoplasm of yeast by placing them in nutrient-deficient media, and autophagy is essential for yeast survival.¹²⁹ This autophagy mechanism also exists in human cells and is closely related to cellular senescence.^{130,131} Mitophagy is a specific autophagic process that removes damaged or excess mitochondria from cells. Much evidence has shown that the level of defective mitochondria in the cell increases during ageing, the integrity of mitochondria is damaged, and faulty mitochondria cannot be cleared, accelerating the cell ageing process.^{132,133} Cells generally clear damaged mitochondria through the mechanism of mitochondrial autophagy,¹²⁰ which can be divided into three pathways: (I) Ubiquitin-dependent PINK1/Parkin-mediated mitochondrial phagocytosis. When mitochondria are damaged, PINK1 stabilizes in the OMM, activating Parkin for subsequent mitochondrial protein ubiquitination. Finally, autophagy receptors such as p62 were recruited to mediate the phagocytosis of mitochondria with the autophagosome membrane interacting with

microtubule-associated protein 1 light chain 3 (MAP1LC3/LC3). The endoplasmic reticulum provides a possible source of autophagosome membranes, in which the autophagic core complexes vacuolar protein sorting 34 (VPS34) and unc-51-like kinase 1 (ULK1) initiate membrane formation. Membrane formation is further mediated by WD repeat domain, phosphoinositide interacting 1 (WIPI1) and WD repeat domain, phosphoinositide interacting 2 (WIPI2), then recombinant autophagy related protein 16 like protein 1 (ATG16L1) and LC3 are recruited, which promotes the formation of autophagosomes. The autophagosome fuses with the acidic lysosome to remove the damaged mitochondria, a step regulated by the synergistic action of the autophagosome and lysosome proteins.^{134,135} (II) Ubiquitin-independent receptor-mediated mitochondrial phagocytosis. Autophagy receptor proteins such as recombinant BCL2/Adenovirus E1B 19 kDa interacting protein 3 (BNIP3), BNIP3-like (NIX), and FUN14 domain containing 1 (FUNDC1) are recruited to the OMM. The receptor proteins interact directly with LC3 to mediate mitochondrial elimination. BNIP3 is an induction of BCL2-interacting protein 3, a mitochondrial outer membrane protein that induces mitochondrial phagocytosis in the adult nervous system and prevents the accumulation of dysfunctional mitochondria in the elderly brain.¹³⁶ (III) Alternative degradation pathway. Cellular pathway of local mitochondrial degradation through partial mitochondrial phagocytosis and mitochondria-derived vesicles to degrade damaged mitochondria and macromolecules.¹³⁷ (Fig. 4). The age-related weakening of mitochondrial autophagy affects removing damaged cellular components and accumulating waste products, including dysfunctional mitochondria in cells. This promotes cell ageing. The mtDNA replication defects, errors, and/or repair failures in senescent cells are the leading causes of

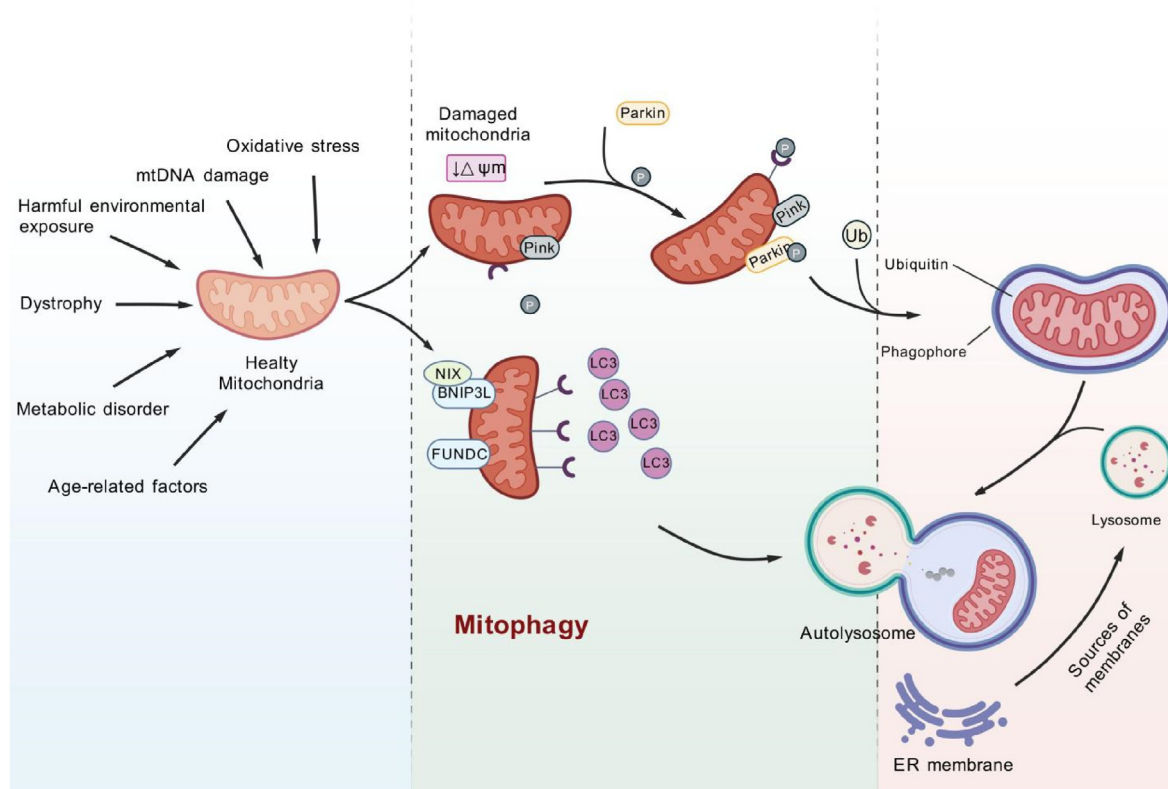


Fig. 4. Mitophagy. Ubiquitin-dependent: PINK/Parkin-mediated mitochondrial phagocytosis: PINK is localized to the outer mitochondrial membrane, recruiting Parkin activation and subsequent ubiquitination of mitochondrial proteins. As an autophagy signal, the autophagy receptor p62 and others are recruited to interact with LC3 and mediate the formation of autophagosomes. Ubiquitin-independent receptor-mediated mitochondrial phagocytosis: BNIP3L and NIX autophagy receptors are recruited in the outer mitochondrial membrane to form autophagosomes by interacting with LC3. In addition, in the ATP supply and hypoxia stress cases, fully activated AMPK α , through phosphorylation of ULK1, promotes VPS34 complex formation and further phosphorylation of the mitochondrial membrane protein FUNDC, mediated by interaction with LC3, facilitating mitochondria and autophagosome integration to complete mitophagy. The endoplasmic reticulum provides a source of autophagosome membranes. We confirm that all figures/images in this article are original and were created by the BioGDP.^{139,140}

mitochondrial biological damage, mitochondrial autophagy decline, and cell bioenergy failure, which promote the permanent cell cycle arrest of cells and accelerate ageing and tissue lesions.¹³⁸ To sum up, mitochondrial homeostasis is necessary to maintain the normal state of cells; that is, the balance of mitochondrial biogenesis and mitochondrial autophagy is conducive to the extension of a healthy life. The failure of damaged mitochondria to be effectively cleared can bring about a series of cellular stress responses that aggravate ageing and age-related diseases, especially neurodegenerative diseases.

3.3. Mitochondrial dynamics

Mitochondria are highly dynamic organelles continuously dividing and fusing to form a reticular state within cells.¹⁴¹ Mitochondria achieve intracellular energy distribution, transfer proton flow, and control steady-state Ca^{2+} concentration through the reticular state, facilitating efficient ATP production and distribution.^{142–144} The inner membrane area of mitochondria in senile cardiomyocytes decreased significantly with ageing, the respiratory capacity decreased, and mitochondria deteriorated gradually.¹⁴⁵ Mitochondrial DNA mutation and accumulation of unfolded or damaged mitochondrial proteins induced by excess ROS exceed a certain threshold to perform mitochondrial dysfunction in aged cells. Mitochondrial Quality Control, a pathway of gene repair and degradation of damaged proteins, mitigates this deterioration at the molecular level. Normal cells had a mitochondrial antioxidant defence system against oxidative damage caused by excess ROS. In addition, mitochondria complete mitochondrial quality control through fission and fusion pathways.¹⁴⁶ The heart is the organ that needs the most energy and is rich in healthy mitochondria, which rarely fuse or fission and remain static in heart muscle cells.¹⁴⁷ The key to cellular mitochondrial quality control is maintaining its network structure's dynamic balance. Two neighbouring mitochondria fuse, and one fissions into two. If this dynamic mitochondrial plasticity is unbalanced, a nutrient-rich environment accompanies a fragmented mitochondrial network, whereas starvation leads to elongated and fused mitochondria.¹⁴⁸

Interestingly, the mitochondrial division is unequal; upon an increase in the impairment within the mitochondria, damaged compartments are segregated through fission and cleared by the mitochondrial autophagy pathway.¹⁴⁸ The primary function of mitochondrial dynamin is to mediate mitochondrial fusion and division. When mitochondrial dynamin cannot function normally, the damaged and dysfunctional mitochondria accumulate and accelerate the occurrence or development of neurodegenerative diseases, muscular dystrophy, sarcopenia, senescence, and metabolic diseases.^{149,150} Regulating mitochondrial fission and fusion to maintain the number of healthy mitochondria in cells is an effective method to treat cardiovascular diseases and promote the function of an ageing heart.¹¹⁵ The abundant OMM protein provides a target for regulating mitochondrial dynamics. OMM protein coupled with continuous fission and fusion. Mitochondrial division separates the damaged mitochondria and eliminates them through mitochondrial autophagy. Mitochondrial fusion mixes slightly damaged mitochondria with healthy ones to repair damage through functional complementarity and to inhibit the abnormal increase in mitochondrial autophagy arising from the accumulation of damaged mitochondria.¹⁵¹ Deletion of some OMM proteins, such as FUNDC1, Mitofusin 1 (MFN1), and Metaxin 2 (MTX2), induces mitochondrial dysfunction, including network fragmentation and oxidative phosphorylation, resulting in cell senescence and decreased proliferation.^{152–154}

Mitochondrial fusion is completed by a two-step process involving simultaneous fusion of OMM and IMM. Currently, it is known that three kinetic protein-related GTPases are involved: MFN1, MFN2, and OPA1 mitochondrial dynamin-like GTPase (OPA1).¹⁵⁵ MFN1 works in mitochondrial tethering, whereas MFN2 operates in a later fusion reaction process.¹⁵⁶ OPA1 is a crucial regulator of mitochondrial intima fusion and crest remodelling.¹⁵⁷ Mitochondria fuse to form tubular or elongated mitochondria to create a network of interconnected mitochondria.

Mitochondrial fission involves three steps: (I) Mark the site of division, and mitochondrial fission factor (MFF) is anchored to OMM as a binding site for dynamin-related protein 1 (DRP1), which is recruited around the mitochondria. (II) The DRP1 dimer and oligomer assemble into a helical superstructure around the labelled fission site. (III) The helical superstructure continuously shrinks and severs OMM and IMM by GTP hydrolysis. Mitochondrial dynamic proteins of 49 (MID49), mitochondrial dynamic proteins of 51 (MID51), and mitochondrial fission 1 protein (FIS1) mediate the aggregation of DRP1 in OMM.¹⁵⁵ DRP1, as the primary regulator driving mitochondrial fission, is regulated by mitochondrial phosphatase PGAM family member 5 (PGAM5) and clustered mitochondria homolog (CLUH). PGAM5 activates DRP1 to participate in mitochondrial division through dephosphorylation. CLUH binds to the mRNA encoding DRP1 receptors MID49 and MFF to up-regulate their expression. Promoting recruitment of DRP1 from cytosol to OMM.^{158,159} Therefore, the dynamic processes of mitochondrial fission and fusion are strictly regulated to maintain the homeostasis and dynamic balance of the mitochondrial network, which is essential for normal cells.¹⁶⁰ Mitochondrial network disruption is an essential manifestation of mitochondrial dysfunction and promotes cell ageing. Therefore, activating mitochondrial division and fusion to restore the integrity of the mitochondrial network is a promising treatment for age-related diseases.¹⁶¹ To target MFN1 fusion activity, a small molecule agonist was developed. Termed S89, it rescued mitochondrial fragmentation and swelling following ischemia/reperfusion injury by interacting with the GTPase domain of MFN1,¹⁶² thus delaying ageing-derived senescence resulting from mitochondrial DNA mutations. Another study identified a peptidomimetic small molecule, MASM7. MASM7 activates MFN2 pro-tethering conformation and enables mitochondrial fusion, resulting in increased membrane potential, mitochondrial respiration, and subsequent ATP production, promising to reduce age-related degenerative metabolic disease.¹⁶³ Targeting mitochondrial dynamics for the prevention and treatment of cellular senescence and age-related diseases is strong.

3.4. Mitochondrial protein quality control

More than 40 proteases in different compartments of mitochondria are collectively known as mitoproteases, which limit the accumulation of short-lived, regulatory proteins within mitochondria, modulate the activity of mitochondrial proteins, and mediate the degradation of damaged proteins. The loss of mitoproteases seriously damages the functional integrity of mitochondria, promotes senescence, and leads to various diseases.^{134,164} Impaired or dysfunctional mitochondrial protease function is associated with ageing and a lot of pathologic conditions, such as neurodegenerative diseases, metabolic syndrome, and cancer. According to their function and location, mitochondrial proteases can be divided into three main categories, including: (I) "Intrinsic mitochondrial proteases", which function mainly in the mitochondrion; (II) "Pseudo-mitotic proteases", which have structural similarity to proteases but lack some critical residues for catalysis, and these can regulate the activity of homologous proteases. (III) "Transient mitotic proteases" are mainly related to apoptosis or autophagy and translocate to mitochondria to perform additional proteolytic activities. The above three groups can be further divided into three subgroups according to the catalytic category: Cys, metalloproteinases, and Ser proteinases.¹⁶⁵ Mitochondrial protein generation requires a complex folding and assembly process to obtain the active state of enzymes. Therefore, endogenous enzymatic components, including multiple chaperones and proteases as interconnected functional networks, are the basis for maintaining mitochondrial protein homeostasis.¹⁶⁶ Moreover, the loss of membrane homeostasis is the leading cause of the degradation of mitochondrial protein quality. Studies have shown that mitochondria-associated endoplasmic reticulum membranes (MAMs) in rats' hearts and skeletal muscles are damaged during ageing. The decrease of highly interconnected proteins with ageing in an ER-mitochondria organization network, including

voltage-dependent anion channel 1 (VDAC1), SAMM50, MIC60, and MTX1, has been identified as a potential factor for age-related MAMs dysfunction.¹⁶⁷ FUNDC1, containing the FUN14 domain, is an OMM protein that mediates the formation of MAMs, whereas increasing MAMs promote angiogenesis and delay the occurrence of age-related neurodegenerative diseases, inflammation, and metabolic dysfunction.¹⁶⁸

Lon proteases degrade misfolded proteins as a significant component of the energy-dependent proteolytic enzyme system in the mitochondrial matrix compartment. In addition, mitochondria also activate the mitochondrial unfolded protein response (UPRmt), which helps misfolded proteins return to regular protein conformation and assists the correct folding of newly synthesized proteins. HDA-1, the *Caenorhabditis elegans* ortholog of mammalian histone deacetylase (HDAC) is required for mitochondrial stress-mediated activation of UPRmt. *C. elegans* responds to mitochondrial stress by activating UPRmt to buffer the mitochondrial folding environment, rewire the metabolic state, and promote innate immunity and lifespan extension.¹¹⁴ Lon protease homolog 1 (LONP1) is a principal mitochondrial protease resident in the matrix. The gradual loss of mammalian LONP1 level with ageing promotes the abnormal accumulation of mitochondrial retention proteins in muscle, inducing the activation of the autophagic lysosomal degradation program of muscle loss and causing muscle loss and weakness.¹⁶⁹ Sustained mitochondrial health depends on coordinated biogenesis and clearance, both regulated by continuously targeting proteins to organelles.¹⁷⁰ This requires a constant supply of mitochondrial protein input. In the mammalian PD model, it was found that the downregulation of the mitochondrial translocase of outer mitochondrial membrane 20 (TOM20) and translocase of inner mitochondrial membrane 23 (TIM23) leads to the inhibition of complex I, which damages the mitochondrial protein input, disrupts the mitochondrial protein balance, and further deteriorates the development of age-related diseases.¹⁷¹ Lon protease deficiency reduces energy production of oxidative phosphorylation and decreases mitochondrial gene expression.¹⁷² ER is a place for protein processing in cells. ER membrane is associated with almost all cell compartments, and the mitochondria-ER contact site (MERCs) may be crucial in developing ageing and age-related diseases. Mitochondrial or ER dysfunction results from oxidative stress, mtDNA mutations, misfolded protein accumulation, and organelle turnover defects.¹⁷³ The ER inositol 1,4,5-trisphosphate receptor, type 2 (ITPR2) calcium-release channel, and calcium fluxes from the ER to the mitochondria are drivers of senescence in human cells. ITPR2 knockout mice showed extended lifespans and reduced age-related phenotypes, possibly due to ITPR2 loss that weakened MERCs and increased MERCs sufficient to trigger premature cell ageing.¹⁷⁴

3.5. Oxidative damage reaction and oxidative phosphorylation activity decreased in senescent cells

Stem cells or progenitor cells are generally considered candidates for promising regenerative applications because of their high proliferation capacity and the potential to differentiate into other cell types^{175,176}; however, stem cells in senescent tissues exhibit the characteristics of cellular senescence and the accumulation of senescent cells with decreased regeneration and differentiation potential is one of the significant factors in the tissue lesions associated with senescent diseases.¹⁷⁷ Mesenchymal stem cells (MSCs) are an emerging strategy for treating many degenerative diseases. However, there is evidence that the function of MSC decreases with age, manifested by an oxidative stress damage state of increased mitochondrial ROS and decreased mitochondrial membrane potential. Compared with the young sample, NADH dehydrogenase (ubiquinone) iron-sulfur protein 6 (NDUFS6) is depressed in aged MSCs.¹⁷⁸ Mitochondrial activation is critical to the differentiation and proliferation of stem cells, and they are also essential cell cycle regulators.¹⁷⁹ Two critical cyclically regulated signaling pathways, the p53 and p38 signaling pathways, control the increase of mitochondrial mass, membrane potential, and cell ROS production. The effects of ROS

on human endometrial-derived MSCs were tested by exposing them to non-lethal doses of hydrogen peroxide, and hydrogen peroxide was found to cause cell senescence through the p53/p21/pRb and p38/MAPKAPK2 pathways. Increasing ROS levels cause most cells to enter irreversible cell cycle arrest.¹⁸⁰

Werner syndrome (WS) is a premature ageing disorder caused by mutations in a RecQ-like DNA helicase. Mice lacking the helicase domain of the Wrn homolog exhibit many phenotypic features of WS, including oxidative stress and metabolic dysfunction, compared to wild-type (WT) animals. Compared with WT mice, the levels of antioxidant proteins such as glutathione (GSH) were reduced. ROS levels in liver tissues were increased, and the difference was more significant in the elderly stage.¹⁸¹ The depletion of NAD⁺ in WS mice showed reduced total ATP synthesis and impaired mitochondrial autophagy. NAD⁺ repletion can effectively prolong the lifespan of WS and delay accelerated ageing by restoring NAD⁺ metabolic profile and improving mitochondrial quality through cell cycle-related factors Cdt1 and ULK1-dependent mitophagy.¹⁸² With the increase of age, the activity of the two central proteolytic systems involved in protein quality control in cells, namely, the autophagy-lysosome system and the ubiquitin-proteasome system (UPS), decreases with ageing. The efficiency of the DNA repair pathway and protein balance pathway is reduced, resulting in the accumulation of ROS, damage to mitochondrial integrity and cell homeostasis, and promotion of cell ageing.⁶⁷ Ageing muscle shows significantly reduced regeneration, and the expression of the "anti-ageing" protein α -Klotho decreases with age, resulting in loss of mitochondrial crest integrity and decreased mitochondrial productivity, which inhibits functional regeneration of aged muscle in vivo.¹⁸³ An age-dependent deterioration in mitochondrial morphology was also shown in PD, induced by the loss of Noto3, a major developmental transcription factor in progressive degeneration of dopaminergic neurons.¹⁸⁴ Therefore, mitochondrial integrity destruction and oxidative stress induce cell senescence, and senescent cells promote ROS production and damage accumulation, further promoting cell senescence. Notably, although free radicals are generally associated with cell damage and inflammation at high levels, they may also protect organisms from more significant subsequent stress at lower levels through an adaptive response called "Mitohormesis".¹⁸⁵

Mitochondria are considered to be the main sites of ROS production. In the process of electron transfer in mitochondrial respiration, the leaking electron and molecular oxygen generate a lot of superoxide anion ($O_2^{\cdot -}$) and hydrogen peroxide (H_2O_2), and some mitochondria are damaged.¹⁸⁶ Excessive ROS can also destroy mitochondrial membrane permeability, leading to the disorder of Ca^{2+} balance inside and outside mitochondria, and Ca^{2+} efflux acts as a second messenger to initiate multiple signaling pathways and promote apoptosis. Mitochondrial calcium uptake family member 3 (MICU3), a regulator of MCU, might maintain normal levels of mitochondrial Ca^{2+} , attenuate oxidative stress and apoptosis, and restore age-related loss of skeletal muscle mass and function.⁶⁶ Prolonged exposure to high levels of ROS results in injury to cellular macromolecules (including proteins essential for oxidative phosphorylation) and mtDNA.¹⁸⁷ Even the mitochondrial dysfunction in peripheral blood mononuclear cells (PBMCs) that shows reduced mitochondrial ATP production, increased proton leakage, and oxidative stress can also give rise to mild cognitive impairment (MCI) in the elderly population.¹⁸⁸ Mitochondria are driven by the electrochemical gradient along the respiratory chain to generate ATP through oxidative phosphorylation (OXPHOS) in an oxygen-dependent manner.¹⁷⁹ The activity of OXPHOS gradually declines during ageing, as demonstrated in fibroblasts from patients with Hutchinson-Guilford progeria syndrome (HGPS).¹⁸⁹ The expression of cytochrome c, complex IV component cytochrome c oxidase subunit I (COXI), and β -ATPase of complex V protein was significantly lower than that of normal cells, resulting in severe damage of OXPHOS. Therefore, ageing is closely related to the reduction of mitochondrial energy production, involving the inhibition of the mitochondrial tricarboxylic acid (TCA) cycle, the inhibition of the glycolytic pathway, and the significant decrease in the activity level of

mitochondrial respiratory complex protein.^{150,190,191} OXPHOS, as the primary production mode of cells, has decreased activity in ageing.

Meanwhile, abnormal mitochondrial function produces excess ROS, which promotes cell ageing.¹¹⁶ Instead, mitochondria have higher activity in cells at earlier developmental stages, and the OXPHOS activity is higher. Mitochondria have efficient ATP production capacity and self-repair ability.¹⁹² Due to oxidative stress damage, excessive ROS cannot be cleared effectively. Ageing can produce large ROS production, destroy protein balance, change genomic stability, and promote cell ageing.¹⁹³ NRF2 is a key transcription factor for the antioxidant response, and its activity decreases gradually with the increase of age.¹⁹⁴ However, the total content of NRF2 in the heart of old and young mice is similar at the non-stress basal level.¹⁹⁵ Therefore, the activity of NRF2 is the key to delaying cell senescence. But Kelch-like ECH-associated protein-1 (Keap1), Glycogen synthase kinase-3 β (GSK-3 β), essential leucine zipper transcription factor 1 (Bach1), and p53 participate in the negative regulatory mechanism of NRF2 to reduce its activity with ageing.¹⁹⁶ Meanwhile, it has been shown that maintaining NRF2 activity can protect cells from the effects of oxidative stress and ageing.^{197,198}

4. Mitochondrial function maintenance and mitochondrial training to delay cellular senescence and treat related diseases

4.1. Supplementation

A variety of natural compounds have been found to enhance mitochondrial function and protect healthy mitochondrial networks, which is expected to slow down the ageing process. Caffeine is gradually favoured by the vast majority of people in daily life. Studies have found that after long-term caffeine consumption, mitochondrial function is improved, autophagy level is increased, antioxidant protein expression is up-regulated, and intestinal atrophy is inhibited in intestinal ageing.¹⁹⁹ Coenzyme Q10 is a highly effective antioxidant that has been shown to have anti-ageing effects. Studies have reported that coenzyme Q10 reversed age-dependent loss of mitochondrial function in reversed epithelial tissues by improving epithelial cells' electron transport chain capacity and antioxidant capacity and increasing ATP production.²⁰⁰ Sulforaphane is a well-known activator of NRF2. By increasing the expression and activity of NRF2 in the renal cortex and decreasing the protein expression of NRF2 repressor Keap1, it can significantly enhance the antioxidant capacity of kidney mitochondria in aged mice and improve kidney injury but does not affect the expression of NRF2 in young mice.²⁰¹ Some peptides and natural antioxidants targeting mitochondria, polyphenols, can prevent retinal nerve ageing and delay vision loss, mainly by protecting mitochondrial function and inhibiting Akt/mTOR activity, to prevent age-related eye diseases and delay ageing.^{202–204} Ageing is characterized by failure to maintain protein balance and mitochondrial homeostasis, accumulation of starch deposits, and mitochondrial dysfunction in skeletal muscle. NAD⁺ supplement nicotinamide riboside (NR) and high-intensity interval training (HIIT) were found to effectively stimulate the mitochondrial imbalance, and UPRmt to restore mitochondrial homeostasis and reduce amyloid accumulation.^{205,206} Lipoic acid is a compound that eliminates free radicals that contribute to accelerated ageing and disease. It has been found that lipoic acid supplementation may be an adjuvant to the treatment of mitochondrial damage and cognitive decline associated with ageing and neurodegenerative diseases.²⁰⁷ Spermidine (SPD), a major mammalian polyamine, can delay cardiac ageing by activating mitochondrial biogenesis. Studies have shown that SPD activates mitochondrial biogenesis through SIRT1-mediated PGC-1 α deacetylation to alleviate cardiac ageing.²⁰⁸ It has been reported that alginate oligosaccharides (AOS) have the potential anti-ageing activity of improving mitochondrial biogenesis, maintaining mitochondrial integrity, and enhancing the adequate clearance of damaged mitochondria. In addition, AOS can also reduce ROS production and oxidative stress state, which further inhibits the destruction of heart mitochondria.²⁰⁹

Endothelial cell senescence is a significant risk factor in cardiovascular diseases. Metformin is an activator of the AMPK pathway, which retards endothelial cell senescence through the upregulation of DOT1, which is a highly conserved methyltransferase of lysine 79 (H3K79) of histone H3 mediated by the deacetylase SIRT1. Increased trimethylation of H3K79 can promote mitochondrial biogenesis and function. Furthermore, a chronic low-dose administration of metformin significantly attenuated vascular ageing and inhibited age-associated atherosclerotic plaque formation in ApoE^{−/−} mice.²¹⁰ Eramitide (SS-31) and Nicotinamide Mononucleotide (NMN) were able to reverse age-related changes in the hearts of mice. In addition, both were effective in upregulating the abundance of electron-transport chain-associated proteins that are depleted by ageing and in deacetylating highly acetylated mitochondrial proteins such as pyruvate dehydrogenase, carnitine palmitoyltransferase 1, and ATPase 50.²¹¹ PGC-1 proteins are master regulators of energy production and expenditure in the cell; senescence of retinal pigment epithelial (RPE) cells may be associated with changes in PGC-1 α function. It was shown that PGC-1 α was involved in antioxidant defence through interaction with Nfe212 (nuclear factor erythroid 2-related factor 2), a transcription factor important in protection against oxidative stress resulting from damage and inflammation.²¹² That study also showed that superoxide dismutase 2 (SOD2), an enzyme clearing an excess of mitochondrial ROS, was activated by PGC-1 α .⁹⁰ Table S1 lists additional anti-ageing strategies, focusing on mitochondrial research.

Testosterone supplementation improved exploratory behaviour, attenuated neuronal degeneration and neuronal loss, and ameliorated mitochondrial dysfunction by enhancing both antioxidative capacity and biogenesis of mitochondria in the aged male rats. Besides, testosterone enhances the function of the mitochondrial complex V in the substantia nigra. It upregulates ATP levels, to some extent, maintains dopaminergic function in the striatum nigra in aged male rats.^{213,214} Substantia nigra provides dopamine for the brain, but there is mitochondrial dysfunction of the substantia nigra in the early stage of ageing and Parkinson's disease. Melatonin is an endogenous free radical scavenger synthesized by neuronal mitochondria and decreased with ageing and neurodegeneration. Inadequate melatonin levels disrupt mitochondrial homeostasis, releasing mtDNA and activating cellular soluble DNA-mediated neuronal inflammatory responses.²¹⁵ (Table S1).

4.2. Lifestyle

In recent years, the function of CR in humans, which can delay ageing and promote health, has been widely discussed. CR has been proven to promote mitochondrial generation in liver tissue and protect mtDNA from damage with the increase of age in mammals, but CR has a weakened effect on improving ageing in older mice (over 32 months of age). However, this only partially limits CR's anti-ageing and improvement of mitochondrial dysfunction.²¹⁶ Ageing-like changes in skeletal muscle can also be ameliorated by proper exercise and intermittent eating throughout life.^{217,218} Diet-related anti-ageing methods have been utilized for anti-ageing, such as CR and intermittent fasting, which have been proven to have sound effects on prolonging ageing, and dietary changes and additives have shown beneficial effects on improving mitochondrial quality and ageing.^{219,220} Table S1 lists additional anti-ageing strategies: focusing on mitochondrial research.

4.3. Exercise

The mechanisms by which endurance exercise improves mitochondrial quality, quantity, and function focus on the following: activation of the PGC-1 α and AMPK signaling pathways to increase healthy mitochondrial biosynthesis and clearance of damaged mitochondria; enhancement of the efficiency of oxidative phosphorylation and antioxidant capacity; and regulation of the expression of mitochondrial fusion and fission proteins to maintain the dynamic balance of the mitochondrial network in response to different intra-/external cellular stresses.

Taking HIIT as an example, HIIT modulates a series of cellular adaptive responses related to mitochondrial biogenesis and mitochondrial autophagy in skeletal muscle by activating the mitogen-activated protein kinases (MAPK) signaling pathway and PGC-1 α and by compensating for the increase in the production of ROS.²²¹ Whereas endurance training increased the expression levels of proteins and mRNAs of mitochondrial growth transcriptional regulators, it also increased the content of mtDNA in skeletal muscle, such as *cAMP*, *PKA*, *AMPK*, *PGC-1 α* , *CREB*, *SIRT1/3*, *TFAM*, and *NRF1/2*.²²² Moreover, prolonged exercise has also been shown to stimulate mitochondrial dilation, resulting in increased mitochondrial ATP production and tolerance to environmental and climatic changes after exercise, but the mechanism of this response remains to be elucidated.²²³ Effective types of exercise and protective measures include aerobic exercise (e.g., brisk walking, swimming, running, etc.), strength training (exercise using equipment or one's body weight), balance and coordination training (e.g., tai chi, yoga), and flexibility training (stretching). For protective measures, warm up and stretch before exercise, avoid over-exercise, arrange exercise time reasonably, use suitable exercise equipment, and reduce sedentary time.^{224,225} (Table S1).

4.4. Transplantation and other

Unlike classical drug therapy, mitochondria of healthy ovarian cells are transferred to abnormal ovarian cells through micromanipulation, which can improve the fertility of low-quality oocytes for age-related infertility in female germ cells.²²⁶ Moreover, the mitochondrial transfer restores mitochondrial function, reduces ROS levels, and activates mitochondrial autophagy to clear damaged mitochondria and improve cell function in different ageing cells.^{227–229} Similarly, healthy mitochondria can also affect damaged mitochondria in the ageing brain, restoring mitochondrial homeostasis and improving cognitive impairment.²³⁰ Mitochondrial density is greatest in photoreceptors, particularly cones that have high energy demands and mediate colour vision. Hence, the retina ages faster than other organs, with a 70% ATP reduction over life and a significant decline in photoreceptor function. Mitochondria have specific light absorbance characteristics influencing their performance. Shinjmar uses 670 nm light to improve photoreceptor performance and measures this psychophysically in those aged 28–72 years. Rod and cone performance declined significantly after approximately 40 years of age. 670 nm light had no impact on younger individuals, but in those around 40 years and older, significant improvements were obtained in colour contrast sensitivity for the blue visual axis (tritan) known to display mitochondrial vulnerability.²³¹ More interventions are listed in Table S1.

5. Summary and outlook

Cell senescence, characterized by permanent stagnation of proliferation, is a response to both endogenous and exogenous stress. As the number of senescent cells increases, different tissues and organs will show various degrees of ageing, and some stem cells will also enter the senescence stage. During ageing, some decreases are observed in various physiological functions such as immune function, hormone levels, brain cells' number and connections with each other, and myocardial contractility. These get worse in neurodegenerative diseases, myocardial infarction, sarcopenia, and metabolic syndrome. SASP secretion amplifies the effects of endogenous cell proliferation stagnation and impairs tissue regeneration in chronic ageing-related diseases. Cellular senescence is a double-edged sword. The beneficial role of senescent cells in embryonic development is prominent. Cellular senescence is helpful for certain vital activities, such as normal development and morphogenesis of embryos, tissue repair, wound healing, and inhibition of tumor growth. However, the accumulation of senescent cells, beyond the body's ability to clear and repair, can induce chronic inflammation and a decline in tissue and organ function, contributing to age-related disease and even a soaring likelihood of cancer.

First of all, with the increase of age, the number of healthy mitochondria decreases in the cell, and some defective mitochondria accumulate in the cell and damage the mitochondrial network, reducing the efficiency of energy generation and destroying intracellular and extracellular signal transmission, which accelerates cell senescence. Secondly, the dynamic changes of mitochondria are controlled by mitochondrial fusion and division for the regular operation of the whole mitochondrial network in the cell. Mitochondria enhance communication between each other and maintain cell homeostasis through the fusion mechanism; damaged mitochondrial parts are eliminated by division, and healthy mitochondrial parts are preserved for efficient cell metabolism. If damaged mitochondria could not be repaired as above, the accumulation of damaged proteins and the loss of critical proteases in mitochondria would aggravate mitochondrial dysfunction, further inhibiting mitochondrial turnover, ineffective fusion, and division, and boosting cell senescence. At the same time, mitochondrial dysfunction would produce excessive ROS in oxidative phosphorylation to produce ATP, damage ageing mitochondrial mtDNA by oxidation, and reduce the quality of mitochondria and energy production. Lots of ROS would also damage other cellular macromolecules and destroy the typical morphology of cells. Thus, maintaining the mitochondrial proteome and homeostasis is highly important in cellular survival and organismal health.²³²

A small amount of mitochondrial dysfunction does not necessarily hurt the cell, as within a specific range, the mitochondria clear these dysfunctional mitochondria through autophagy.¹⁸⁵ What is the threshold level for accumulation of mitochondrial dysfunction, and how does accumulation of mitochondrial DNA replication damage reach that threshold level remain to be investigated. It remains to be studied how a certain degree of mitochondrial DNA replication damage can be controlled at a threshold level. The function of mitochondria is greatly affected by external pressure, which makes mitochondria an excellent model against the odds.

Healthy mitochondria are vital for maintaining the normal function of cells, so it is interesting to find appropriate methods to improve mitochondrial quality to reduce cell senescence or improve the function of senescent cells. Research on mitochondria as a target for anti-cellular senescence has focused on two main areas: to improve and/or restore mitochondrial function and to improve the number of mitochondria. Some substances help improve mitochondrial function by promoting mitochondrial autophagy, increasing mitochondrial biogenesis, regulating mitochondrial fusion and division, restoring mitochondrial protein homeostasis, improving mitochondrial energy production efficiency, and improving the oxidative defense system. Mitochondria are considered highly plastic organelles. This plasticity enables the mitochondria to undergo morphological and functional changes in response to cellular demands.²³³ Mitochondria exhibit different morphologies, such as enlargement, elongation, fragmentation, or aggregation of the mitochondrial network to regulate energy metabolism in the cell and adapt to internal damage and external environmental stressors.^{234,235}

Cellular senescence is triggered by a variety of factors, including mitochondrial dysfunction. However, there is no clear causal relationship between cellular senescence and mitochondrial dysfunction in the biological ageing process, and together they characterize senescence. With age, the mitochondrial antioxidant defence system gradually weakens, leading to excessive accumulation of ROS. These ROS damage the mitochondria's DNA, proteins, and lipids and affect other components of the cell, ultimately leading to cellular dysfunction and senescence. Although the removal of senescent cells is simple and effective, the issue of specificity is more challenging to achieve, whereas the protection and activation of mitochondria can occur at all stages of life and can be achieved in a variety of ways, so targeting mitochondria is considered to be more effective in improving the function of senescent cells and in mitigating the onset and progression of age-related diseases. Given the highly dynamic and plastic nature of the mitochondrial network, is it possible to continuously train mitochondria through chronically low levels of stimulation, such as exercise, dietary changes, nutrient intake, or active

substance and drug interventions? More research is needed to make mitochondria more powerful, more efficient at self-healing and waste removal, and with higher critical levels of resistance to environmental degradation. Mitochondrial function and regulatory mechanisms are complex, and there are many unknowns and challenges in the research of anti-ageing and ageing-related diseases related to restoring damaged mitochondrial function.

CRediT authorship contribution statement

Gan Gao: Writing – original draft, Software, Methodology, Investigation. **Zhihui Xie:** Resources. **Hongliang Huang:** Writing – review & editing, Validation, Supervision, Resources, Project administration, Funding acquisition, Conceptualization.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.jhip.2025.04.001>.

Abbreviations

MQC	Mitochondrial Quality Control	CIP1	cyclin-dependent kinase (CDK)-inhibitor 1
SASP	senescence-associated secretory phenotype	ROS	reactive oxygen species
OS	oxidative stress	MMP	mitochondrial membrane potential
IL-6	Interleukin-6	TNF-α	tumor necrosis factor-α
HUVECs	human umbilical vein endothelial cells	CNS	central nervous system
AD	Alzheimer's disease	PD	Parkinson's disease
ALS	Amyotrophic Lateral Sclerosis	HD	Huntington's disease
Aβ	β-amyloid protein	Caspase-3	cysteiny l aspartate specific proteinase
DAMP	damage-associated molecular pattern	UTD	Urinary Tract Dysfunction
IPF	Idiopathic Pulmonary Fibrosis	CKD	Chronic Kidney disease
FOXO4	forkhead box O 4	COPD	Chronic Obstructive Pulmonary disease
ECFCs	endothelial colony-forming cells	AT II	type II alveolar epithelial cells
AMD	age-related macular degeneration	IMM	inner mitochondrial membrane
OMM	outer mitochondrial membrane	IBM	inner boundary membrane
T2DM	Type 2 diabetes mellitus	PGC-1α	peroxisome proliferator-activated receptor gamma coactivator-1α
AMPK	adenosine 5'-monophosphate (AMP)-activated protein kinase	POLG	mitochondrial DNA polymerase γ
TFAM	mitochondrial transcription factor A	ERRα	estrogen-related receptor α
SIRT1	silent information regulator 1	CR	caloric restriction
NRF2	NF-E2 p45-related factor 2	LC3	microtubule-associated protein 1 light chain 3
VPS34	vacuolar protein sorting 34	ULK1	unc-51-like kinase 1
WIP1	WD repeat domain, phosphoinositide interacting 1	ATG16L1	autophagy related protein 16 like protein 1
BNIP3	recombinant BCL2/Adenovirus E1B 19 kDa interacting protein 3	MFN1	Mitofusin1
MTX2	Metaxin 2	DRP1	dynamin-related protein 1
MID49	Mitochondrial dynamic proteins of 49	FIS1	mitochondrial fission 1 protein
PGAM5	mitochondrial phosphatase PGAM family member 5	CLUH	clustered mitochondria homolog
VDAC1	voltage dependent anion channel 1	UPRmt	mitochondrial unfolded protein response
LONP1	Lon protease homolog 1	TOM20	outer mitochondrial membrane 20
TIM23	inner mitochondrial membrane 23	MERCS	mitochondria-ER contact site
ITPR2	ER inositol 1,4,5-trisphosphate receptor, type 2	MSCs	Mesenchymal stem cells
NDUFS6	NADH dehydrogenase (ubiquinone) iron-sulfur protein 6	WS	Werner syndrome
UPS	ubiquitin-proteasome system	H ₂ O ₂	hydrogen peroxide
MICU3	Mitochondrial calcium uptake family member 3	PBMCs	peripheral blood mononuclear cells
MCI	mild cognitive impairment	OXPHOS	oxidative phosphorylation
HGPS	Hutchinson-Guilford progeria syndrome	TCA	tricarboxylic acid cycle
Keap1	Kelch-like ECH-associated protein-1	GSK-3β	Glycogen synthase kinase-3β
Bach1	essential leucine zipper transcription factor 1	NR	nicotinamide riboside
HIIT	high-intensity interval training	SPD	Spermidine
AOS	alginate oligosaccharides	H3K79	highly conserved methyltransferase of lysine 79
NMN	Nicotinamide Mononucleotide	SOD2	superoxide dismutase 2
MAPK	mitogen-activated protein kinases		

References

1. Khosla S, Farr JN, Tchkonja T, et al. The role of cellular senescence in ageing and endocrine disease. *Nat Rev Endocrinol.* 2020;16(5):263–275.

2. Amorim JA, Coppotelli G, Rolo AP, et al. Mitochondrial and metabolic dysfunction in ageing and age-related diseases. *Nat Rev Endocrinol.* 2022;18(4):243–258.

3. Florian JB, Stephen WGT. Mitochondria as multifaceted regulators of cell death. *Nat Rev Mol Cell Biol.* 2019;21(2):85–100.

4. Matthew Yoke Wui N, Timothy W, Anne S. Quality control of the mitochondrion. *Dev Cell.* 2021;56(7):881–905.

5. Eva-Maria E, Olga Z, Luisa K, et al. Sensing, signaling and surviving mitochondrial stress. *Cell Mol Life Sci.* 2021;78(16):5925–5951.

6. Sturm G, Karan KR, Monzel AS, et al. OxPhos defects cause hypermetabolism and reduce lifespan in cells and in patients with mitochondrial diseases. *Commun Biol.* 2023;6(1):22.

7. Rossignol RMM, Mazat JP, Letellier T. Threshold effect and tissue specificity. Implication for mitochondrial cytopathies. *J Biol Chem.* 1999;33426–33432.

8. Lopez-Otin C, Blasco MA, Partridge L, et al. Hallmarks of aging: an expanding universe. *Cell*. 2023;186(2):243–278.
9. Memme JM, Erlich AT, Phukan G, et al. Exercise and mitochondrial health. *J Physiol*. 2021;599(3):803–817.
10. Merry BJ. Oxidative stress and mitochondrial function with aging—the effects of calorie restriction. *Aging Cell*. 2004;3(1):7–12.
11. Siegel MP, Kruse SE, Percival JM, et al. Mitochondrial-targeted peptide rapidly improves mitochondrial energetics and skeletal muscle performance in aged mice. *Aging Cell*. 2013;12(5):763–771.
12. Leonardo AD. *DNA Damage Triggers a Prolonged P53-dependent G1 Arrest and Long-Term Induction of Cip1 in Normal Human Fibroblasts*. Cold Spring Harbor Laboratory Press; 1994:2540–2551.
13. Gire V, Dulic V. Senescence from G2 arrest, revisited. *Cell Cycle*. 2015;14(3):297–304.
14. Childs BG, Baker DJ, Kirkland JL, et al. Senescence and apoptosis: dueling or complementary cell fates? *EMBO Rep*. 2014;15(11):1139–1153.
15. Loiza N, Demaria M. Cellular senescence and tumor promotion: is aging the key? *Biochim Biophys Acta*. 2016;1865(2):155–167.
16. Helmut S. Oxidative stress: concept and some practical aspects. *Antioxidants (Basel)*. 2020;9(9):852.
17. Afsar B, Afsar RE. Hypertension and cellular senescence. *Biogerontology*. 2023;24(4):457–478.
18. Chandrika C, Mark DS, Aleksandr EV, et al. Increased mitochondrial NADPH oxidase 4 (NOX4) expression in aging is a causative factor in aortic stiffening. *Redox Biol*. 2019;26:101288.
19. Farhan R, Claudia CP, Larisa E, et al. Effects of aging on cardiac oxidative stress and transcriptional changes in pathways of reactive oxygen species generation and clearance. *J Am Heart Assoc*. 2021;10(16):e019948.
20. Yusuf O, Erkan T, Sinan D, et al. Ageing-associated increase in SGLT2 disrupts mitochondrial/sarcoplasmic reticulum Ca(2+) homeostasis and promotes cardiac dysfunction. *J Cell Mol Med*. 2020;24(15):8567–8578.
21. Arielys M, Jason K. Keeping the beat against time: mitochondrial fitness in the aging heart. *Front Aging*. 2022;3:951417.
22. Hiroe T, Merry LL. Extracellular matrix roles in cardiorenal fibrosis: potential therapeutic targets for CVD and CKD in the elderly. *Pharmacol Ther*. 2018;193:99–120.
23. He C, Yunxia L, Hongbo M, et al. Protective mechanism of humanin against oxidative stress in aging-related cardiovascular diseases. *Front Endocrinol (Lausanne)*. 2021;12(0):683151.
24. Jeremy T, Michael M, Qun C. Targeting ER stress and calpain activation to reverse age-dependent mitochondrial damage in the heart. *Mech Ageing Dev*. 2020;192(0):111380.
25. Daniel M, Cécile B, Matthew EP, et al. The exceptional longevity of the naked mole-rat may be explained by mitochondrial antioxidant defenses. *Aging Cell*. 2019;18(3):e12916.
26. Alessia M, Alessandro M, Martin G, et al. Mitochondrial epigenetics in aging and cardiovascular diseases. *Front Cardiovasc Med*. 2023;10(1):11.
27. Biyi C, Nastaran D, Hsiang-Chun L, et al. Mitochondrial oxidative stress mediates bradyarrhythmia in leigh syndrome mitochondrial disease mice. *Antioxidants (Basel)*. 2023;12(5):1001.
28. Ronny L, Inez FY, Hanna G, et al. Low dose of β -carotene regulates inflammation, reduces caspase signaling, and correlates with autophagy activation in cardiomyoblast cell lines. *Med Sci Monit Basic Res*. 2020;26:e928648.
29. Wenjing F, Jianya L, Shan W, et al. Alginate oligosaccharide alleviates D-galactose-induced cardiac ageing via regulating myocardial mitochondria function and integrity in mice. *J Cell Mol Med*. 2021;25(15):7157–7168.
30. Cesari M, Penninx BW, Newman AB, et al. Inflammatory markers and cardiovascular disease (the health, aging and body composition [health ABC] study). *Am J Cardiol*. 2003;92(5):522–528.
31. Bokov A, Chaudhuri A, Richardson D. The role of oxidative damage and stress in aging. *Mech Ageing Dev*. 2004;125(10–11):811–826.
32. Rock KL, Kono H. The inflammatory response to cell death. *Annu Rev Pathol*. 2008;3:99–126.
33. Ding YN, Wang HY, Chen HZ, et al. Targeting senescent cells for vascular aging and related diseases. *J Mol Cell Cardiol*. 2021;162(0):43–52.
34. Roland S. Senotherapy: growing old and staying young? *Pflügers Archiv*. 2017;469(9):1051–1059.
35. Matthew H, Yoko I, Tae-Won K, et al. NOTCH1 mediates a switch between two distinct secretomes during senescence. *Nat Cell Biol*. 2016;18(9):979–992.
36. Yu S, Jean-Philippe C, Eric WFL. Cellular senescence: the sought or the unwanted? *Trends Mol Med*. 2018;24(10):871–885.
37. Mattson MP, Arumugam TV. Hallmarks of brain aging: adaptive and pathological modification by metabolic states. *Cell Metab*. 2018;27(6):1176–1199.
38. Satoh A, Imai SI, Guarente L. The brain, sirtuins, and ageing. *Nat Rev Neurosci*. 2017;18(6):362–374.
39. Hou Y, Dan X, Babbar M, et al. Ageing as a risk factor for neurodegenerative disease. *Nat Rev Neurol*. 2019;15(10):565–581.
40. Andra IT, Carl WC. Emerging roles of oxidative stress in brain aging and Alzheimer's disease. *Neurobiol Aging*. 2021;107:86–95.
41. Nicole GR, D Allan B. Altered metabolism in alzheimer disease brain: role of oxidative stress. *Antioxidants Redox Signal*. 2021;36(16–18):1289–1305.
42. Heather MW, Russell HS. Mitochondrial links between brain aging and Alzheimer's disease. *Transl Neurodegener*. 2021;10(1):33.
43. Bocheva G, Bakalov D, Iliev P, et al. The vital role of melatonin and its metabolites in the neuroprotection and retardation of brain aging. *Int J Mol Sci*. 2024;25(10):5122.
44. Antignano I, Liu Y, Offermann N, et al. Aging microglia. *Cell Mol Life Sci : CMLS*. 2023;80(5):126.
45. Benjamin GT, Dominic JH, Kay LD. Oxidative stress in the aging substantia nigra and the etiology of Parkinson's disease. *Aging Cell*. 2019;18(6):e13031.
46. Badanjak K, Fixemer S, Smajić S, et al. The contribution of microglia to neuroinflammation in Parkinson's disease. *Int J Mol Sci*. 2021;22(9):203.
47. Asghar M, Odeh A, Fattahi AJ, et al. Mitochondrial biogenesis, telomere length and cellular senescence in Parkinson's disease and Lewy body dementia. *Sci Rep*. 2022;12(1):17578.
48. Picca A, Mankowski RT, Burman JL, et al. Mitochondrial quality control mechanisms as molecular targets in cardiac ageing. *Nat Rev Cardiol*. 2018;15(9):543–554.
49. Leng F, Edison P. Neuroinflammation and microglial activation in Alzheimer disease: where do we go from here? *Nat Rev Neurol*. 2021;17(3):157–172.
50. Chen HL, Liao F, Lin TN, et al. Galectins and neuroinflammation. *Adv Neurobiol*. 2014;9:517–542.
51. Odrodnik M, Evans SA, Fielder E, et al. Whole-body senescent cell clearance alleviates age-related brain inflammation and cognitive impairment in mice. *Aging Cell*. 2021;20(2):e13296.
52. Curtis E, Litwic A, Cooper C, et al. Determinants of muscle and bone aging. *J Cell Physiol*. 2015;230(11):2618–2625.
53. Gomarasca M, Banfi G, Lombardi G. *Myokines: The Endocrine Coupling of Skeletal Muscle and Bone [M]*. 2020:155–218.
54. Malvandi AM, Shahba S, Mehrzad J, et al. Metabolic disruption by naturally occurring mycotoxins in circulation: a focus on vascular and bone homeostasis dysfunction. *Front Nutr*. 2022;9:915681.
55. Moiseeva V, Cisneros A, Sica V, et al. Senescence atlas reveals an aged-like inflamed niche that blunts muscle regeneration. *Nature*. 2022;613(7942):169–178.
56. Gerosa L, Malvandi AM, Malavolta M, et al. Exploring cellular senescence in the musculoskeletal system: any insights for biomarkers discovery? *Ageing Res Rev*. 2023;88:101943.
57. Baht GS, Silkstone D, Vi L, et al. Exposure to a youthful circulation rejuvenates bone repair through modulation of β -catenin. *Nat Commun*. 2015;6(1):7131.
58. Baker D, Childs B, Durik M, et al. Naturally occurring p16(Ink4a)-positive cells shorten healthy lifespan. *Nature*. 2016;530(7589):184–189.
59. Farr J, Xu M, Weivoda M, et al. Corrigendum: targeting cellular senescence prevents age-related bone loss in mice. *Nat Med*. 2017;23(11):1384.
60. Fuhrmann-Stroisnigg H, Ling Y, Zhao J, et al. Identification of HSP90 inhibitors as a novel class of senolytics. *Nat Commun*. 2017;8(1):422.
61. Chen M, Wang Y, Deng S, et al. Skeletal muscle oxidative stress and inflammation in aging: focus on antioxidant and anti-inflammatory therapy. *Front Cell Dev Biol*. 2022;10:964130.
62. Salmon AB, Richardson A, Pérez VI. Update on the oxidative stress theory of aging: does oxidative stress play a role in aging or healthy aging? *Free Radic Biol Med*. 2010;48(5):642–655.
63. Gilles G, Martin P, Richard G, et al. Role of peroxisome proliferator-activated receptor gamma coactivator 1- α (PGC-1 α) in denervation-induced atrophy in aged muscle: facts and hypotheses. *Longev Heal*. 2014;2(1).
64. Gouspillou G, Sgaroto N, Kapchinsky S, et al. Increased sensitivity to mitochondrial permeability transition and myonuclear translocation of endonuclease G in atrophied muscle of physically active older humans. *FASEB J*. 2013;28(4):1621–1633.
65. Drummond MJ, Addison O, Brunker L, et al. Downregulation of E3 ubiquitin ligases and mitophagy-related genes in skeletal muscle of physically inactive, frail older women: a cross-sectional comparison. *J Gerontol Series A: Biol Sci Med Sci*. 2014;69(8):1040–1048.
66. Yang YF, Yang W, Liao ZY, et al. MICU3 regulates mitochondrial Ca(2+)-dependent antioxidant response in skeletal muscle aging. *Cell Death Dis*. 2021;12(12):1115.
67. Lopez-Otin C, Blasco MA, Partridge L, et al. The hallmarks of aging. *Cell*. 2013;153(6):1194–1217.
68. Kim IH, Kisseleva T, Brenner DA. Aging and liver disease. *Curr Opin Gastroenterol*. 2015;31(3):184–191.
69. Odrodnik M, Miwa S, Tchkonja T, et al. Cellular senescence drives age-dependent hepatic steatosis. *Nat Commun*. 2017;8:15691.
70. Koliaki C, Szendroedi J, Kaul K, et al. Adaptation of hepatic mitochondrial function in humans with non-alcoholic fatty liver is lost in steatohepatitis. *Cell Metab*. 2015;21(5):739–746.
71. Stahl EC, Haschak MJ, Popovic B, et al. Macrophages in the aging liver and age-related liver disease. *Front Immunol*. 2018;9:2795.
72. Niemann J, John C, Schröder S, et al. An mtDNA mutation accelerates liver aging by interfering with the ROS response and mitochondrial life cycle. *Free Radic Biol Med*. 2017;102:174–187.
73. Passos JF, Nelson G, Wang C, et al. Feedback between p21 and reactive oxygen production is necessary for cell senescence. *Mol Syst Biol*. 2010;6(1):347.
74. Almalki WH, Almurji SS. Aging, ROS, and cellular senescence: a trilogy in the progression of liver fibrosis. *Biogerontology*. 2024;26(1):10.
75. Teresa L, Emily R, Donald DF, et al. Mitochondrial dysfunction and cellular senescence due to aging contributes to prostatic fibrosis. *Innovation in Aging*. 2020;4(supplment_1):133–134.
76. Aghali A, Koloko Ngassie ML, Pabelick CM, et al. Cellular senescence in aging lungs and diseases. *Cells*. 2022;11(11):1781.
77. Woldhuis RR, Heijink IH, van den Berge M, et al. COPD-derived fibroblasts secrete higher levels of senescence-associated secretory phenotype proteins. *Thorax*. 2021;76(5):508–511.
78. Schafer MJ, White TA, Iijima K, et al. Cellular senescence mediates fibrotic pulmonary disease. *Nat Commun*. 2017;8:14532.

79. Yao C, Guan X, Carraro G, et al. Senescence of alveolar type 2 cells drives progressive pulmonary fibrosis. *Am J Respir Crit Care Med*. 2021;203(6):707–717.
80. Schmitt R, Melk A. Molecular mechanisms of renal aging. *Kidney Int*. 2017;92(3):569–579.
81. Schmitt R, Melk A. New insights on molecular mechanisms of renal aging. *Am J Transplant*: official journal of the American Society of Transplantation and the American Society of Transplant Surgeons. 2012;12(11):2892–2900.
82. Zhang Y, Yu C, Li X. Kidney aging and chronic kidney disease. *Int J Mol Sci*. 2024;25(12):6585.
83. Franzin R, Stasi A, Ranieri E, et al. Targeting premature renal aging: from molecular mechanisms of cellular senescence to senolytic trials. *Front Pharmacol*. 2021;12:630419.
84. Allison S. Ageing: targeting senescence-associated tissue damage. *Nat Rev Nephrol*. 2017;13(6):319.
85. Velarde MC, Demaria M. Targeting senescent cells: possible implications for delaying skin aging: a mini-review. *Gerontology*. 2016;62(5):513–518.
86. Barbara R, Nicolò CB, Carlo C. Interplay between keratinocytes and fibroblasts: a systematic review providing a new angle for understanding skin fibrotic disorders. *Front Immunol*. 2020;11(0):648.
87. Ho CY, Dreesen O. Faces of cellular senescence in skin aging. *Mech Ageing Dev*. 2021;198:111525.
88. López-Luppo M, Catita J, Ramos D, et al. Cellular senescence is associated with human retinal microaneurysm formation during aging. *Investigat Ophthalmol; Visual Sci*. 2017;58(7):2832–2842.
89. Guo L, Wang N, Chen J, et al. Cellular senescence and glaucoma. *Exp Gerontol*. 2025;202:112718.
90. Kaarniranta K, Kajdane J, Morawiec J, et al. PGC-1 α protects RPE cells of the aging retina against oxidative stress-induced degeneration through the regulation of senescence and mitochondrial quality control. The significance for AMD pathogenesis. *Int J Mol Sci*. 2018;19(8):2317.
91. Palmer AK, Xu M, Zhu Y, et al. Targeting senescent cells alleviates obesity-induced metabolic dysfunction. *Aging Cell*. 2019;18(3):e12950.
92. Shenghui H, Norman ES. Senescence in health and disease. *Cell*. 2017;169(6):1000–1011.
93. Janko NZ. The twilight of immunity: emerging concepts in aging of the immune system. *Nat Immunol*. 2017;19(1):10–19.
94. van Deursen JM. The role of senescent cells in ageing. *Nature*. 2014;509(7501):439–446.
95. Single-cell profiling of the immune landscape across the human lifespan. *Nat Immunol*. 2025;26(2):172–173.
96. Bennett GC, Matej D, Darren JB, et al. Cellular senescence in aging and age-related disease: from mechanisms to therapy. *Nat Med*. 2015;21(12):1424–1435.
97. Yasar Arfat TK, Robert AJS. Anti-ageing antibodies revive the immune system. *Nature*. 2024;628(8006):43–45.
98. Olivia VB, Roel PHDM, Arne NA. Enhancing immunity during ageing by targeting interactions within the tissue environment. *Nat Rev Drug Discov*. 2025;24:300–312.
99. Marta P, Akang Leonard B, Victoria MS, et al. Targeted clearance of senescent cells using an antibody-drug conjugate against a specific membrane marker. *Sci Rep*. 2021;11(1):20358.
100. Peisu Z, Yuki K, Ioannis G, et al. Senolytic therapy alleviates A β -associated oligodendrocyte progenitor cell senescence and cognitive deficits in an Alzheimer's disease model. *Nat Neurosci*. 2019;22(5):719–728.
101. Tovar J. Mitochondrial remnant organelles of Giardia function in iron-sulphur protein maturation. *Nature PublishingGroup*. 2003;426(6963):172–176.
102. van der Giezen M. Hydrogenosomes and mitosomes: conservation and evolution of functions. *J Eukaryot Microbiol*. 2009;56(3):221–231.
103. Palade G. An electron microscope study of the mitochondrial structure. *J Histochem Cytochem*. 1953;1(4):188.
104. Frey TG. Insight into mitochondrial structure and function from electron tomography. *Biochim Biophys Acta*. 2002;1555(1–3):196–203.
105. Roger AJ, Munoz-Gomez SA, Kamikawa R. The origin and diversification of mitochondria. *Curr Biol*. 2017;27(21):R1177–R1192.
106. De Stefani D, Raffaello A, Teardo E, et al. A forty-kilodalton protein of the inner membrane is the mitochondrial calcium uniporter. *Nature*. 2011;476(7360):336–340.
107. Baughman JM, Perocchi F, Girgis HS, et al. Integrative genomics identifies MCU as an essential component of the mitochondrial calcium uniporter. *Nature*. 2011;476(7360):341–345.
108. Friedman JR, Nunnari J. Mitochondrial form and function. *Nature*. 2014;505(7483):335–343.
109. Sasaki A. Development of diabetes in Japanese subjects with impaired glucose tolerance a seven year follow up study. *Diabetologia*. 1982;22(3):154–157.
110. Gm R. Pathophysiology of insulin resistance in human disease. *Physiol Rev*. 1995;75(3):473–486.
111. Rowe JOHNW. Characterization of the insulin resistance of aging. *Lancet*. 1989;71(6):1581–1587.
112. Petersen KF, Befroy D, Dufour S, et al. Mitochondrial dysfunction in the elderly: possible role in insulin resistance. *Science*. 2003;300(5622):1140–1142.
113. Sun N, Youle RJ, Finkel T. The mitochondrial basis of aging. *Mol Cell*. 2016;61(5):654–666.
114. Shao LW, Peng Q, Dong M, et al. Histone deacetylase HDA-1 modulates mitochondrial stress response and longevity. *Nat Commun*. 2020;11(1):4639.
115. Picca A, Calvani R, Coelho-Junior HJ, et al. Mitochondrial dysfunction, oxidative stress, and neuroinflammation: intertwined roads to neurodegeneration. *Antioxidants*. 2020;9(8):647.
116. Kubben N, Misteli T. Shared molecular and cellular mechanisms of premature ageing and ageing-associated diseases. *Nat Rev Mol Cell Biol*. 2017;18(10):595–609.
117. Golpich M, Amini E, Mohamed Z, et al. Mitochondrial dysfunction and biogenesis in neurodegenerative diseases: pathogenesis and treatment. *CNS Neurosci Ther*. 2017;23(1):5–22.
118. Scholpa NE, Lynn MK, Corum D, et al. 5-HT1F receptor-mediated mitochondrial biogenesis for the treatment of Parkinson's disease. *Br J Pharmacol*. 2018;175(2):348–358.
119. Van Laar VS, Arnold B, Howlett EH, et al. Evidence for compartmentalized axonal mitochondrial biogenesis: mitochondrial DNA replication increases in distal axons as an early response to Parkinson's disease-relevant stress. *J Neurosci*. 2018;38(34):7505–7515.
120. Lopez-Lluch G, Irueta PM, Navas P, et al. Mitochondrial biogenesis and healthy aging. *Exp Gerontol*. 2008;43(9):813–819.
121. Rowe GC, Jiang A, Arany Z. PGC-1 coactivators in cardiac development and disease. *Circ Res*. 2010;107(7):825–838.
122. Fernandez-Marcos PJ, Auwerx J. Regulation of PGC-1 α , a nodal regulator of mitochondrial biogenesis. *Am J Clin Nutr*. 2011;93(4):884S–890S.
123. Khadije Hasanpour BA. Lida moradi the effect of eight weeks of moderate-intensity continuous training and high intensity interval training along with *Citrus aurantium* L on some cardiac mitochondrial biogenesis markers in the heart tissue of elderly rats. *North Khorasan Univ Med Sci*. 2022;10(2):1.
124. Lee G, Uddin MJ, Kim Y, et al. PGC-1 α , a potential therapeutic target against kidney aging. *Aging Cell*. 2019;18(5):e12994.
125. Nisoli SF E, Tonello C, Cozzi V, Palomba L, Fiorani M. Mitochondrial biogenesis by NO yields functionally active mitochondria in mammals. *Proc Natl Acad Sci U S A*. 2004;101(47):16507–16512.
126. Canto C, Auwerx J. PGC-1 α , SIRT1 and AMPK, an energy sensing network that controls energy expenditure. *Curr Opin Lipidol*. 2009;20(2):98–105.
127. Shimizu Y, Polavarapu R, Eskla KL, et al. Hydrogen sulfide regulates cardiac mitochondrial biogenesis via the activation of AMPK. *J Mol Cell Cardiol*. 2018;116:29–40.
128. Dinkova-Kostova AT, Abramov AY. The emerging role of Nrf2 in mitochondrial function. *Free Radic Biol Med*. 2015;88(Pt B):179–188.
129. Kazuhiko Takeshige MB, Tsuboi Shigeru, Noda Takeshi, Ohsumi Yoshinori. Autophagy in yeast demonstrated with proteinase-deficient mutants and conditions for its induction. *J Cell Biol*. 1992;119(2):301–311.
130. Tran M, Reddy PH. Defective autophagy and mitophagy in aging and Alzheimer's disease. *Front Neurosci*. 2020;14:612757.
131. Korolchuk VI, Miwa S, Carroll B, et al. Mitochondria in cell senescence: is mitophagy the weakest link? *EBioMedicine*. 2017;21:7–13.
132. Youle RJ. Mitochondria-Striking a balance between host and endosymbiont. *Science*. 2019;365(6454).
133. Kauppila TES, Kauppila JHK, Larsson NG. Mammalian mitochondria and aging: an update. *Cell Metab*. 2017;25(1):57–71.
134. Palikaras K, Lionaki E, Tavernarakis N. Mechanisms of mitophagy in cellular homeostasis, physiology and pathology. *Nat Cell Biol*. 2018;20(9):1013–1022.
135. Pickles S, Vigie P, Youle RJ. Mitophagy and quality control mechanisms in mitochondrial maintenance. *Curr Biol*. 2018;28(4):R170–R185.
136. Schmid ET, Pyo JH, Walker DW. Neuronal induction of BNIP3-mediated mitophagy slows systemic aging in Drosophila. *Nat Aging*. 2022;2(6):494–507.
137. Bakula D, Scheibye-Knudsen M. Mitophagy: mitophagy in aging and disease. *Front Cell Dev Biol*. 2020;8:239.
138. Vasileiou PVS, Evangelou K, Vlasik K, et al. Mitochondrial homeostasis and cellular senescence. *Cells*. 2019;8(7):686.
139. Ramaccini D, Montoya-Urbe V, Aan FJ, et al. Mitochondrial function and dysfunction in dilated cardiomyopathy. *Front Cell Dev Biol*. 2021;8:624216.
140. Shuai J, Huiqin L, Luowanyue Z, et al. Generic Diagramming Platform (GDP): a comprehensive database of high-quality biomedical graphics. *Nucleic Acids Res*. 2024;53(D1):D1670–D1676.
141. Cao YL, Meng S, Chen Y, et al. MFN1 structures reveal nucleotide-triggered dimerization critical for mitochondrial fusion. *Nature*. 2017;542(7641):372–376.
142. chas FJL, Mazat JP. Mitochondria are excitable organelles capable of generating and conveying electrical and calcium signals. *Cell*. 1997;89(7):1145–1153.
143. Glancy B, Hartnell LM, Malide D, et al. Mitochondrial reticulum for cellular energy distribution in muscle. *Nature*. 2015;523(7562):617–620.
144. Amchenkova AABL, Chentsov YS, Skulachev VP, et al. Coupling membranes as energy-transmitting cables. I. Filamentous mitochondria in fibroblasts and mitochondrial clusters in cardiomyocytes. *J Cell Biol*. 1988;107(2):481–495.
145. El'darov Ch M, Vays VB, Vangeli IM, et al. Morphometric examination of mitochondrial ultrastructure in aging cardiomyocytes. *Biochemistry (Mosc)*. 2015;80(5):604–609.
146. Fischer F, Hamann A, Osiewacz HD. Mitochondrial quality control: an integrated network of pathways. *Trends Biochem Sci*. 2012;37(7):284–292.
147. Dorn 2nd GW. Evolving concepts of mitochondrial dynamics. *Annu Rev Physiol*. 2019;81:1–17.
148. Liu YJ, McIntyre RL, Janssens GE, et al. Mitochondrial fission and fusion: a dynamic role in aging and potential target for age-related disease. *Mech Ageing Dev*. 2020;186:111212.
149. Sebastián D, Zorzano A. Mitochondrial dynamics and metabolic homeostasis. *Curr Opin Physiol*. 2018;3:34–40.
150. Tian C, Kim YJ, Hali S, et al. Suppressed expression of LDHB promotes age-related hearing loss via aerobic glycolysis. *Cell Death Dis*. 2020;11(5):375.
151. Xian H, Liou YC. Functions of outer mitochondrial membrane proteins: mediating the crosstalk between mitochondrial dynamics and mitophagy. *Cell Death Differ*. 2021;28(3):827–842.

152. Elouej S, Harhour K, Le Mao M, et al. Loss of MTX2 causes mandibuloacral dysplasia and links mitochondrial dysfunction to altered nuclear morphology. *Nat Commun.* 2020;11(1):4589.
153. Yu T, Fox RJ, Burwell LS, et al. Regulation of mitochondrial fission and apoptosis by the mitochondrial outer membrane protein hFis1. *J Cell Sci.* 2005;118(Pt 18): 4141–4151.
154. Liu L, Feng D, Chen G, et al. Mitochondrial outer-membrane protein FUNDC1 mediates hypoxia-induced mitophagy in mammalian cells. *Nat Cell Biol.* 2012; 14(2):177–185.
155. Al Ojaimi M, Salah A, El-Hattab AW. Mitochondrial fission and fusion: molecular mechanisms, biological functions, and related disorders. *Membranes (Basel).* 2022; 12(9):893.
156. Filadi R, Greotti E, Pizzo P. Highlighting the endoplasmic reticulum-mitochondria connection: focus on Mitofusin 2. *Pharmacol Res.* 2018;128:42–51.
157. Belenguer P, Pellegrini L. The dynamin GTPase OPA1: more than mitochondria? *Biochim Biophys Acta.* 2013;1833(1):176–183.
158. Yu B, Ma J, Li J, et al. Mitochondrial phosphatase PGAM5 modulates cellular senescence by regulating mitochondrial dynamics. *Nat Commun.* 2020;11(1):2549.
159. Yang H, Sibilla C, Liu R, et al. Clueless/CLUH regulates mitochondrial fission by promoting recruitment of Drp1 to mitochondria. *Nat Commun.* 2022;13(1):1582.
160. Sabouny R, Shutt TE. Reciprocal regulation of mitochondrial fission and fusion. *Trends Biochem Sci.* 2020;45(7):564–577.
161. Whitley BN, Engelhart EA, Hoppins S. Mitochondrial dynamics and their potential as a therapeutic target. *Mitochondrion.* 2019;49:269–283.
162. Guo Y, Zhang H, Yan C, et al. Small molecule agonist of mitochondrial fusion repairs mitochondrial dysfunction. *Nat Chem Biol.* 2023;19(4):468–477.
163. Zacharioudakis E, Agianian B, Kumar Mv V, et al. Modulating mitofusins to control mitochondrial function and signaling. *Nat Commun.* 2022;13(1):3775.
164. Deshwal S, Fiedler KU, Langer T. Mitochondrial proteases: multifaceted regulators of mitochondrial plasticity. *Annu Rev Biochem.* 2020;89:501–528.
165. Quiros PM, Langer T, Lopez-Otin C. New roles for mitochondrial proteases in health, ageing and disease. *Nat Rev Mol Cell Biol.* 2015;16(6):345–359.
166. Voos W. Chaperone-protease networks in mitochondrial protein homeostasis. *Biochim Biophys Acta.* 2013;1833(2):388–399.
167. Lu X, Gong Y, Hu W, et al. Ultrastructural and proteomic profiling of mitochondria-associated endoplasmic reticulum membranes reveal aging signatures in striated muscle. *Cell Death Dis.* 2022;13(4):296.
168. Wang C, Dai X, Wu S, et al. FUNDC1-dependent mitochondria-associated endoplasmic reticulum membranes are involved in angiogenesis and neovascularization. *Nat Commun.* 2021;12(1):2616.
169. Xu Z, Fu T, Guo Q, et al. Disuse-associated loss of the protease LONP1 in muscle impairs mitochondrial function and causes reduced skeletal muscle mass and strength. *Nat Commun.* 2022;13(1):894.
170. Lionaki E, Gkikas I, Daskalaki I, et al. Mitochondrial protein import determines lifespan through metabolic reprogramming and de novo serine biosynthesis. *Nat Commun.* 2022;13(1):651.
171. Franco-Iborra S, Cuadros T, Parent A, et al. Defective mitochondrial protein import contributes to complex I-induced mitochondrial dysfunction and neurodegeneration in Parkinson's disease. *Cell Death Dis.* 2018;9(11):1122.
172. Voos W, Pollecker K. The mitochondrial lon protease: novel functions off the beaten track? *Biomolecules.* 2020;10(2):253.
173. Moltedo O, Remondelli P, Amodio G. The mitochondria-endoplasmic reticulum contacts and their critical role in aging and age-associated diseases. *Front Cell Dev Biol.* 2019;7:172.
174. Ziegler DV, Vindrieux D, Goehrig D, et al. Calcium channel ITPR2 and mitochondria-ER contacts promote cellular senescence and aging. *Nat Commun.* 2021;12(1):720.
175. Edward J, Caterson LJN, Danielson Keith G, Tuan Rocky S. Human marrow-derived mesenchymal progenitor cells isolation, culture expansion, and analysis of differentiation. *Mol Biotechnol.* 2002;20(3):245–256.
176. Tuan RS, Boland G, Tuli R. Adult mesenchymal stem cells and cell-based tissue engineering. *Arthritis Res Ther.* 2003;5(1):32–45.
177. Boyette LB, Tuan RS. Adult stem cells and diseases of aging. *J Clin Med.* 2014;3(1): 88–134.
178. Zhang Y, Guo L, Han S, et al. Adult mesenchymal stem cell ageing interplays with depressed mitochondrial Ndufs6. *Cell Death Dis.* 2020;11(12):1075.
179. Lane RK, Hilsabeck T, Rea SL. The role of mitochondrial dysfunction in age-related diseases. *Biochim Biophys Acta.* 2015;1847(11):1387–1400.
180. Borodkina A. Interaction between ROS dependent DNA damage mitochondria and p38 MAPK underlies senescence of human adult stem cells. *Aging.* 2014;6(6):481.
181. Massip L, Garand C, Paquet ER, et al. Vitamin C restores healthy aging in a mouse model for Werner syndrome. *FASEB J.* 2010;24(1):158–172.
182. Fang EF, Hou Y, Lautrup S, et al. NAD(+) augmentation restores mitophagy and limits accelerated aging in Werner syndrome. *Nat Commun.* 2019;10(1):5284.
183. Sahu A, Mamiya H, Shinde SN, et al. Age-related declines in alpha-Klotho drive progenitor cell mitochondrial dysfunction and impaired muscle regeneration. *Nat Commun.* 2018;9(1):4859.
184. Miozzo F, Valencia-Alarcon EP, Stickley L, et al. Maintenance of mitochondrial integrity in midbrain dopaminergic neurons governed by a conserved developmental transcription factor. *Nat Commun.* 2022;13(1):1426.
185. Yun J, Finkel T. Mitohormesis. *Cell Metab.* 2014;19(5):757–766.
186. Jiang T, Sun Q, Chen S. Oxidative stress: a major pathogenesis and potential therapeutic target of antioxidative agents in Parkinson's disease and Alzheimer's disease. *Prog Neurobiol.* 2016;147:1–19.
187. de Souza-Pinto NC, Wilson 3rd DM, Stevnsner TV, et al. Mitochondrial DNA, base excision repair and neurodegeneration. *DNA Repair (Amst).* 2008;7(7):1098–1109.
188. Apaijai N, Sriwichaiin S, Phrommintikul A, et al. Cognitive impairment is associated with mitochondrial dysfunction in peripheral blood mononuclear cells of elderly population. *Sci Rep.* 2020;10(1):21400.
189. Rivera-Torres J, Acin-Perez R, Cabezas-Sanchez P, et al. Identification of mitochondrial dysfunction in Hutchinson-Gilford progeria syndrome through use of stable isotope labeling with amino acids in cell culture. *J Proteonomics.* 2013;91: 466–477.
190. Kang W, Harada Y, Yamatoya K, et al. Extra-mitochondrial citrate synthase initiates calcium oscillation and suppresses age-dependent sperm dysfunction. *Lab Invest.* 2020;100(4):583–595.
191. Andreux PA, van Diemen MPJ, Heezen MR, et al. Mitochondrial function is impaired in the skeletal muscle of pre-frail elderly. *Sci Rep.* 2018;8(1):8548.
192. Zhao Z, Yu Z, Hou Y, et al. Improvement of cognitive and motor performance with mitotherapy in aged mice. *Int J Biol Sci.* 2020;16(5):849–858.
193. Sohal RS. Mechanisms of aging: an appraisal of the oxidative stress hypothesis. *Free Radical Biol Med.* 2002;33(5):575–586.
194. Schmidlin CJ, Dodson MB, Madhavan L, et al. Redox regulation by NRF2 in aging and disease. *Free Radic Biol Med.* 2019;134:702–707.
195. Silva-Palacios A, Ostolga-Chavarria M, Buelna-Chontal M, et al. 3-NP-induced Huntington's-like disease impairs Nrf2 activation without loss of cardiac function in aged rats. *Exp Gerontol.* 2017;96:89–98.
196. Silva-Palacios A, Ostolga-Chavarria M, Zazueta C, et al. Nrf2: molecular and epigenetic regulation during aging. *Ageing Res Rev.* 2018;47:31–40.
197. Kloska D, Kopacz A, Piechota-Polanczyk A, et al. Nrf2 in aging - focus on the cardiovascular system. *Vasc Pharmacol.* 2019;112:42–53.
198. Qiu L, Liu X, Xia H, et al. Downregulation of P300/CBP-associated factor protects from vascular aging via Nrf2 signal pathway activation. *Int J Mol Sci.* 2022;23(20): 12574.
199. Min H, Youn E, Shim YH. Long-term caffeine intake exerts protective effects on intestinal aging by regulating vitellogenesis and mitochondrial function in an aged Caenorhabditis elegans model. *Nutrients.* 2021;13(8):2517.
200. Schniethshauer D, Gebhard D, Bergemann J. Age-dependent loss of mitochondrial function in epithelial tissue can be reversed by coenzyme Q10. *J Aging Res.* 2018; 6354680.
201. Mohammad RS, Lokhandwala MF, Banday AA. Age-related mitochondrial impairment and renal injury is ameliorated by sulforaphane via activation of transcription factor NRF2. *Antioxidants (Basel).* 2022;11(1):156.
202. Wang N, Luo ZW, Jin M, et al. Exploration of age-related mitochondrial dysfunction and the antiaging effects of resveratrol in zebrafish retina. *Aging.* 2019;11(10): 3117–3137.
203. Alam NM, Douglas RM, Prusky GT. Treatment of age-related visual impairment with a peptide acting on mitochondria. *Dis Model Mech.* 2022;15(3):dmm048256.
204. Kim SJ, Miller B, Kumagai H, et al. Mitochondrial-derived peptides in aging and age-related diseases. *Geroscience.* 2021;43(3):1113–1121.
205. Cordeiro AV, Peruca GF, Braga RR, et al. High-intensity exercise training induces mitochondrial imbalance and activates the mitochondrial unfolded protein response in the skeletal muscle of aged mice. *Geroscience.* 2021;43(3):1513–1518.
206. Romani M, Sorrentino V, Oh CM, et al. NAD(+) boosting reduces age-associated amyloidosis and restores mitochondrial homeostasis in muscle. *Cell Rep.* 2021; 34(3):108660.
207. Molz P, de Freitas BS, Uberti VH, et al. Effects of lipoic acid supplementation on age- and iron-induced memory impairment, mitochondrial DNA damage and antioxidant responses. *Eur J Nutr.* 2021;60(7):3679–3690.
208. Wang JY, Wang J, Wu FX, et al. Spermidine alleviates cardiac aging by improving mitochondrial biogenesis and function. *aging.* 2019;12(1):650–671.
209. Feng W, Liu J, Wang S, et al. Alginate oligosaccharide alleviates D-galactose-induced cardiac aging via regulating myocardial mitochondrial function and integrity in mice. *J Cell Mol Med.* 2021;25(15):7157–7168.
210. Karnewar S, Neeli PK, Panuganti D, et al. Metformin regulates mitochondrial biogenesis and senescence through AMPK mediated H3K79 methylation: relevance in age-associated vascular dysfunction. *Biochim Biophys Acta, Mol Basis Dis.* 2018; 1864(4 Pt A):1115–1128.
211. Whitson JA, Johnson R, Wang L, et al. Age-related disruption of the proteome and acetylome in mouse hearts is associated with loss of function and attenuated by elamipretide (SS-31) and nicotinamide mononucleotide (NMN) treatment. *Geroscience.* 2022;44(3):1621–1639.
212. Cherry AD, Suliman HB, Bartz RR, et al. Peroxisome proliferator-activated receptor γ Co-activator 1- α as a critical Co-activator of the murine hepatic oxidative stress response and mitochondrial biogenesis in Staphylococcus aureus sepsis. *J Biol Chem.* 2014;289(1):41–52.
213. Yan WS, Kang YX, Zhang GL, et al. Testosterone ameliorates age-related brain mitochondrial dysfunction. *Aging.* 2021;13(12):16229–16247.
214. Tianyun Zhang YW, Kang Yunxiao, Wang Li, et al. Testosterone enhances mitochondrial complex V function in the substantia nigra of aged male rats. *Aging.* 2020;12(11):10398–10414.
215. Jauhari A, Baranov SV, Suofu Y, et al. Melatonin inhibits cytosolic mitochondrial DNA-induced neuroinflammatory signaling in accelerated aging and neurodegeneration. *J Clin Invest.* 2020;130(6):3124–3136.
216. Chimienti G, Picca A, Fracasso F, et al. The age-sensitive efficacy of calorie restriction on mitochondrial biogenesis and mtDNA damage in rat liver. *Int J Mol Sci.* 2021;22(4):1665.
217. Lettieri-Barbato D, Cannata SM, Casagrande V, et al. Time-controlled fasting prevents aging-like mitochondrial changes induced by persistent dietary fat overload in skeletal muscle. *PLoS One.* 2018;13(5):e0195912.

218. Harper C, Gopalan V, Goh J. Exercise rescues mitochondrial coupling in aged skeletal muscle: a comparison of different modalities in preventing sarcopenia. *J Transl Med.* 2021;19(1):71.
219. Villa-Bellosta R. Dietary magnesium supplementation improves lifespan in a mouse model of progeria. *EMBO Mol Med.* 2020;12(10):e12423.
220. Pathak SJ, Baar K. Ketogenic diets and mitochondrial function: benefits for aging but not for athletes. *Exerc Sport Sci Rev.* 2022;44(1):22–37.
221. Guan Y, Drake J, Yan Z. Exercise-induced mitophagy in skeletal muscle and heart. *Exerc Sport Sci Rev.* 2019;47(3):151–156.
222. Jia D, Tian Z, Wang R. Exercise mitigates age-related metabolic diseases by improving mitochondrial dysfunction. *Ageing Res Rev.* 2023;91:102087.
223. Boushel RL, Carsten Qvortrup, Klaus, Sahlin Kent. Mitochondrial plasticity with exercise training and extreme environments. *Exerc Sport Sci Rev.* 2014;42(4):169–174.
224. Ma L, Li K, Wei W, et al. Exercise protects aged mice against coronary endothelial senescence via FUNDC1-dependent mitophagy. *Redox Biol.* 2023;62:102693.
225. Liu L, Kim S, Buckley M, et al. Exercise reprograms the inflammatory landscape of multiple stem cell compartments during mammalian aging. *Cell Stem Cell.* 2023;30(5):689–705.e684.
226. Mobarak H, Heidarpour M, Tsai PJ, et al. Autologous mitochondrial microinjection; a strategy to improve the oocyte quality and subsequent reproductive outcome during aging. *Cell Biosci.* 2019;9:95.
227. Machihara K, Kageyama S, Oki S, et al. Lotus germ extract rejuvenates aging fibroblasts via restoration of disrupted proteostasis by the induction of autophagy. *Aging.* 2022;14(19):7662–7691.
228. Wang HH, Sun YN, Qu TQ, et al. Nobiletin prevents D-galactose-induced C2C12 cell aging by improving mitochondrial function. *Int J Mol Sci.* 2022;23(19):11963.
229. Visalli G, Ferlazzo N, Facciola A, et al. Ex vivo evaluation of the effects of a white grape juice extract on lymphocytic mitochondrial functions. *Nat Prod Res.* 2018;34(4):580–584.
230. Feng Z. Declining p53 function in the aging process a possible mechanism for the increased tumor incidence in older populations. *Proc Natl Acad Sci U S A.* 2007;104(42):16633–16638.
231. Shinhmar H, Grewal M, Sivaprasad S, et al. Optically improved mitochondrial function redeems aged human visual decline. *J Gerontol: Series A.* 2020;75(9):e49–e52.
232. Eldeeb MA, Thomas RA, Ragheb MA, et al. Mitochondrial quality control in health and in Parkinson's disease. *Physiol Rev.* 2022;102(4):1721–1755.
233. Bahat A, Gross A. Mitochondrial plasticity in cell fate regulation. *J Biol Chem.* 2019;294(38):13852–13863.
234. Mahalingam S, Cheviron ZA, Storz JF, et al. Chronic cold exposure induces mitochondrial plasticity in deer mice native to high altitudes. *J Physiol.* 2020;598(23):5411–5426.
235. Huang Y, Si X, Shao M, et al. Rewiring mitochondrial metabolism to counteract exhaustion of CAR-T cells. *J Hematol Oncol.* 2022;15(1):38.